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(21) International Application Number: PCT/CA92/00246 (22) International Filing Date: 9 June 1992 (09.06.92) (30) Priority data: <table border="0" style="width: 100%;"><tr><td style="width: 33%;">714,161</td><td style="width: 33%;">10 June 1991 (10.06.91)</td><td style="width: 33%;">US</td></tr><tr><td>771,007</td><td>2 October 1991 (02.10.91)</td><td>US</td></tr><tr><td>887,746</td><td>22 May 1992 (22.05.92)</td><td>US</td></tr></table> (71) Applicant: ALBERTA RESEARCH COUNCIL [CA/CA]; 250 Karl Clark Road, P.O. Box 8330, Postal Station F, Edmonton, Alberta T6H 5X2 (CA). (72) Inventors: VENOT, Andre, P. ; 10968-75 Avenue, Edmon- ton, Alberta T6G 0G9 (CA). NIKRAD, Pandurang, V. ; 3619 105th Street, Edmonton, Alberta T6J 2K4 (CA). KASHEM, Mohammed, A. ; 17D Twin Terrace, Edmon- ton, Alberta T6K 1V4 (CA).		714,161	10 June 1991 (10.06.91)	US	771,007	2 October 1991 (02.10.91)	US	887,746	22 May 1992 (22.05.92)	US	(74) Agent: MITCHELL, Richard, J.; Marks & Clerk, 55 Met- calfe Street, Suite 1380, P.O. Box 957, Station B, Ottawa, Ontario K1P 5S7 (CA). (81) Designated States: AT (European patent), BE (European patent), CA, CH (European patent), DE (European pa- tent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), GR (Euro- pean patent), IT (European patent), JP, LU (European patent), MC (European patent), NL (European patent), SE (European patent). Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
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(54) Title: MODIFIED SIALYL LEWIS ^x COMPOUNDS (57) Abstract <p>The present invention is drawn to methods for the synthesis of sialyl Lewis^x derivatives modified at the C-2 and/or C-6 position of GlcNAc employing chemoenzymatic synthesis. The derivatives find use in the treatment and prevention of diseases.</p>											

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MODIFIED SIALYL LEWIS^x COMPOUNDS

BACKGROUND OF THE INVENTION

1. Field of the Invention

5 The present invention is directed to novel sialyl
Lewis^x analogues, pharmaceutical compositions
containing such analogues, methods for their
preparation and methods for their use.

2. References

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All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference in their entirety to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference in its entirety.

10 3. State of the Art

Carbohydrates and/or oligosaccharides are present on a variety of natural and pathological glycoconjugates¹. Of particular interest are carbohydrates and oligosaccharides containing sialic acid residues particularly at the non-reducing sugar terminus³. Such sialic acid terminated carbohydrates and oligosaccharides are present in a number of products which have been implicated in a wide range of biological phenomena based, in part, on the concept of recognition signals carried by the carbohydrate structures and by their binding to specific ligands.

Specifically, such sialic acid terminated carbohydrates and oligosaccharides are believed to be receptors for the binding of toxins⁴, pathogenic agents such as viruses⁵, and are recognition sites for a variety of lectins, particularly those involved in cellular adhesion^{6,7}, etc. Sialylated and sialylated/fucosylated oligosaccharide structures relating to blood group determinants, including sialyl Lewis^x, have also been shown to possess in vivo immunomodulating and tolerogenic properties in mammals². In this regard, the modified sialyl Lewis^x compounds described herein also possess immunomodulating and tolerogenic properties.

Additionally, the presence of certain sialyl terminated oligosaccharides in tumor-related antigens is documented in the art⁸ and, in general, the structures of the oligosaccharides present on such antigens have been modified in some way from normal oligosaccharides so as to lead to the expression of tumor related antigens⁸. The prospect of passive immunotherapy with monoclonal antibodies directed against some sialylated tumor-associated antigens, such as the gangliosides GD₂, GD₃, and GM₂, in patients with melanoma is being investigated^{9,10}. However, most tumor-associated antigens are unable to lead to the production of tumor specific antibodies which would either inhibit or prevent the growth of such tumors. Without being limited to any theory, it is believed that this is due to the absence of real tumor specific antigens and that the structure of such antigens cross-reacts with that of similar structures expressed in a restricted number of normal tissues. In addition, carbohydrate antigens are generally not believed to lead to a T-cell mediated immune response that is expected to play a role in active immunity¹¹. However, some recent studies indicate that, in some cases, tumor-associated carbohydrate antigens may stimulate anticancer T-cell immunity^{12,13} or the production of cytotoxic antibodies¹⁴.

In view of the general inability of carbohydrate tumor-related antigens to produce cytotoxic tumor specific antibodies, it has been proposed to chemically modify naturally occurring weak antigens so as to improve their antigenicity¹⁵. In this regard, methods for chemical modification of specific groups on sialylated carbohydrate tumor-related antigens have been reported.

Much of the focus of chemical modifications to non- or weakly-immunogenic naturally occurring antigens has been to derivatize the sialic acid

residue of sialylated carbohydrate tumor-related antigens. Specifically, the art has reported that some structural modifications present on naturally occurring sialic acids renders the corresponding oligosaccharides immunogenic in selected hosts^{16,17,18,19,20,30}.

Recent work with artificial antigens indicates that while chemically modified sialosides (melanoma associated glycolipid antigens) are antigenic in humans, the antibodies generated in humans by these modified sialosides do not cross-react with the natural substance.⁴⁸ On the other hand, when injected into mice, chemically modified sialylated antigens produce antibodies which do cross-react with the natural substance. Accordingly, cross-reacting monoclonal or polyclonal antibodies generated in mice would serve as a basis for either a diagnostic assay for determining the presence and/or amount of the natural substance in a human host or as a basis for antibody therapy for a disease condition in which the natural substance is attacked by the antibodies which can optionally be coupled to a therapeutic agent.

In regard to the above, Venot et al.²¹ disclose a facile chemical/enzymatic synthesis of sialylated oligosaccharides containing modified sialyl groups at the non-reducing terminus of the oligosaccharide so as to produce a number of sialylated oligosaccharide structures having modifications to the sialic acid residue.

Notwithstanding the advantages of this synthetic method especially as they apply to sialyl Lewis^x compounds modified in the sialyl unit, it would be further advantageous to modify both the galactose and/or the N-acetylglucosamine saccharide units in sialyl Lewis^x so as to provide a variety of structures useful both in modulating an immune response and in preparing artificial antigens for the purpose of preparing antibodies thereto. From a practical point

of view, it would be beneficial to prepare such analogues by a chemo/enzymatic approach where the sialic acid and fucose units are added to the modified $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ type II structure by appropriate sialyltransferases and fucosyltransferases respectively.

In the case of preparing sialyl Lewis^x and analogues thereof, appropriate sialyltransferases include the known $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc}$ $\alpha(2\rightarrow3)$ sialyltransferase from rat liver²⁴ which transfers sialic acid to the 3 position of galactose to form an $\alpha(2\rightarrow3)$ linkage. Appropriate fucosyltransferases include the known $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc}$ $\alpha(1\rightarrow3/4)$ fucosyltransferase which is readily obtained from human milk^{22,27,28} and the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ $\alpha(1\rightarrow3)$ fucosyltransferase which is also found in human serum and is co-recovered with the $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc}$ $\alpha(1\rightarrow3/4)$ fucosyltransferase. A recombinant form of $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc}$ $\alpha(1\rightarrow3/4)$ fucosyltransferase is also available^{25,26}.

In this regard, sialyl Lewis^x structures have heretofore been prepared by a combined chemical/enzymatic approach which involves sequentially sialylating and fucosylating a $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ type II structure with appropriate glycosyltransferases^{22,23}. Specifically, sialylation of the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ type II structure involves contacting this structure with CMP-Neu5Ac in the presence of an appropriate sialyltransferase so as to place the Neu5Ac group at the 3 position of the galactose so as to form an $\alpha(2\rightarrow3)$ linkage which results in the formation of $\alpha\text{Neu5Ac}(2\rightarrow3)\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$. Fucosylation is then accomplished by contacting this compound with GDP-fucose in the presence of an appropriate fucosyltransferase so as to place a fucose group at the 4-position of the GlcNAc unit to form an $\alpha(1\rightarrow3)$

linkage which results in the formation of α Neu5Ac(2 \rightarrow 3) β Gal(1 \rightarrow 4)[α Fuc(1 \rightarrow 3)] β GlcNAc (i.e., sialyl Lewis^x).

5 In regard to the above, the use of sialyltrans-
ferases and fucosyltransferases provides for the most
facile method for the synthesis of sialyl Lewis^x
because, under these conditions, sialylation and
fucosylation do not require protection/deprotection of
10 reactive moieties at other sites of the
oligosaccharide structure as is common in chemical
synthesis. Moreover, sialyltransferases readily form
the α (2 \rightarrow 3) linkage which are otherwise difficult to
form in high yield with anomeric specificity.

In this regard, the art recognizes that certain
15 modifications can be tolerated by the
 β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase on the 2
and 3 positions of the galactose structures of
 β Gal(1 \rightarrow 4) β GlcNAc^{22,29} but that the presence of a Neu5Ac
or deoxy group at the 6-position of the galactose
20 cannot be tolerated by this enzyme. The art also
recognized that the compound resulting from
replacement of the 2-Nac group on GlcNAc unit of
 β Gal(1 \rightarrow 4) β GlcNAc disaccharide with an equatorial
hydroxyl group is a good acceptor for this enzyme
25 whereas the compound resulting from replacement of the
Nac group on GlcNAc with an axial hydroxyl group is
not an acceptor for this enzyme.^{28,29} In any event, the
art is deficient as to what, if any, other
modifications to the GlcNAc structure of the
30 β Gal(1 \rightarrow 4) β GlcNAc disaccharide or the
 α Neu5Ac(2 \rightarrow 3) β Gal(1 \rightarrow 4) β GlcNAc trisaccharide would be
tolerated by the β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyl-
transferase.

The art also recognizes that the β Gal(1 \rightarrow 4) β GlcNAc
35 α (1 \rightarrow 3)fucosyltransferase tolerates modification of the
2- and 3-positions of galactose unit but does not
accept lactose [β Gal(1 \rightarrow 4) β Glc] as a substrate²⁹.

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Preliminary data also indicates that this fucosyltransferase can transfer L-fucose to type II acceptors substituted by a sulfate group at the 2', 3', 6' position of the terminal galactose and the 6-position of the GlcNAc⁶⁷.

Similarly, while the art has heretofore disclosed that $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(2\rightarrow3)\text{sialyltransferase}$ will not transfer Neu5Ac to the $\alpha(2\rightarrow3)$ position of the galactose unit of $\beta\text{Gal}(1\rightarrow4)[\alpha\text{Fuc}(1\rightarrow3)]\beta\text{GlcNAc-structure}^{31}$ but will transfer Neu5Ac to the $\alpha(2\rightarrow3)$ position of the galactose unit of $\beta\text{Gal}(1\rightarrow4)[\text{saccharide}(1\rightarrow6)]\beta\text{GlcNAc-structure}^{89}$, it is not known what other modifications, if any, on the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ type II structure would be tolerated by $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(2\rightarrow3)\text{sialyltransferase}$.

This uncertainty made it difficult to provide any rationale method for modifying the galactose and/or N-acetylglucosamine units of the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ type II structure with the expectation of sequentially sialylating and fucosylating this structure by using the $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(2\rightarrow3)\text{sialyltransferase}$ and then using the $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(1\rightarrow3/4)\text{fucosyltransferase}$ or any other available and acceptable transferase.

In view of the above, it would be particularly beneficial to develop modified sialyl Lewis^x structures which would possess immunomodulating and tolerogenic properties as well as which could be used as antigenic determinants on artificial antigens so to prepare monoclonal and polyclonal antibodies. It would be further beneficial if these modified sialyl Lewis^x structures could be readily prepared by sequentially sialylating and fucosylating a modified $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ structure with an appropriate sialyltransferase and an appropriate fucosyltransferase.

SUMMARY OF THE INVENTION

The present invention is directed in part to the discovery that the modified sialyl Lewis^x compounds described herein are useful in both modulating a cell-mediated immune response to an antigen in vivo while providing tolerance to later challenges from that antigen as well as in preparing artificial antigens to produce antibodies directed to these structures. In this regard, the monoclonal or polyclonal antibodies raised against these artificial antigens (especially in mice) are useful in diagnostic assays for determining the presence of sialyl Lewis^x structures in human hosts. Additionally, it is contemplated that these antibodies would be useful in antibody therapy for a disease condition wherein the antibody would attach to sialyl Lewis^x type antigenic structures on diseased particles (e.g., cancer cells).

The present invention is directed to the further discovery that the modifications made to the galactose and/or N-acetylglucosamine units of sialyl Lewis^x are compatible with specified sialyl- and fucosyltransferases so that the sialyl and fucosyl residues on these compounds are readily prepared by sequential enzymatic methods.

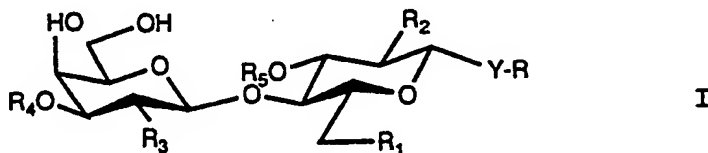
Specifically, it has now been discovered that the β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)sialyltransferase can accept extensive modifications to the 2 and 6 positions of the N-acetylglucosamine unit as well as some modification of the 2 position of the galactose unit of β Gal(1 \rightarrow 4) β GlcNAc type II structures and still readily transfer sialic acid to the 3-position of the galactose to form an α (2 \rightarrow 3) linkage on the galactose saccharide. Similarly, it has now been discovered that β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase can accept extensive modifications to the 6 position and some modification to the 2 position of the

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N-acetylglucosamine unit as well as non-oxygen containing modification at the 2-position of the galactose saccharide of the β Gal(1 \rightarrow 4) β GlcNAc type II structure and still readily transfer L-fucose to form
 5 an α (1 \rightarrow 3) fucose residue on the N-acetylglucosamine structure.

In view of these discoveries, we concluded that the overlap of modifications permitted by the β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)sialyl- and the
 10 β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyl-transferases allowed for modifications at the 2 and 6 positions of the N-acetylglucosamine and the 2 position of the galactose units of β Gal(1 \rightarrow 4) β GlcNAc type II structures and still permit sequential sialylation and
 15 fucosylation by the respective transferases.

Accordingly, in one of its composition aspects, this invention is directed to compounds which are suitable for use in modulating a cell-mediated immune response to an antigen in a mammal which compounds are
 20 represented by Formula I:



where R is selected from the group consisting of hydrogen, a saccharide, an oligosaccharide, or an aglycon having at least 1 carbon atom;

25 R_1 is selected from the group consisting of hydrogen, $-\text{NH}_2$, $-\text{N}_3$, $-\text{NHSO}_3\text{H}$, $-\text{NR}_6\text{C}(\text{O})\text{R}_6$, $-\text{N}=\text{C}(\text{R}_7)_2$, $-\text{NHCH}(\text{R}_7)_2$, $-\text{N}(\text{R}_8)_2$, $-\text{O}(\text{C}(\text{O}))_p\text{R}_9$, $-\text{SR}_8$, fluoro, chloro, bromo, and sulfate,

wherein R_6 is selected from the group consisting
 30 of hydrogen,
 alkyl of from 1 to 4 carbon atoms optionally substituted with 1 or more substituents selected from

the group consisting of hydroxy, chloro, bromo, alkoxy
of from 1 to 4 carbon atoms, phenyl, and phenyl
substituted with 1 to 3 substituents selected from the
group consisting of hydroxy, alkyl of from 1 to 4
5 carbon atoms, alkoxy of from 1 to 4 carbon atoms,
chloro, bromo, and sulfate,
an amino acid or polypeptidyl residue,
-OR₁₀ wherein R₁₀ is alkyl of from 1 to 4
carbon atoms, or alkyl of from 2 to 4 carbon atoms
10 substituted with a hydroxyl group, and
-NR₁₁R₁₂ wherein R₁₁ and R₁₂ are independently
selected from the group consisting of hydrogen and
alkyl of from 1 to 4 carbon atoms,
each R₇ is independently selected from the
15 group consisting of hydrogen and alkyl of from 1 to 4
carbon atoms,
R₈ is independently selected from the group
consisting of hydrogen and alkyl of from 1 to 4 carbon
atoms,
20 R₉ is selected from the group consisting of
hydrogen, alkyl of from 1 to 4 carbon atoms optionally
substituted with 1 or more substituents selected from
the group consisting of hydroxy, chloro, bromo, alkoxy
of from 1 to 4 carbon atoms, phenyl, and phenyl
25 substituted with 1 to 3 substituents selected from the
group consisting of hydroxy, alkyl of from 1 to 4
carbon atoms, alkoxy of from 1 to 4 carbon atoms,
chloro, bromo, and sulfate,
and p is an integer equal to 0 or 1;
30 R₂ is selected from the group consisting of
hydrogen, -N₃, -NH₂, -NHSO₃H, -NR₁₅C(O)R₁₃, -N=C(R₁₄)₂,
-NHCH(R₁₄)₂, -N(R₁₅)₂, -O(C(O))_qR₁₆, fluoro, chloro, bromo
and sulfate,
wherein R₁₃ is selected from the group
35 consisting of hydrogen,
alkyl of from 1 to 4 carbon atoms optionally
substituted with 1 or more substituents selected from

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the group consisting of hydroxy, chloro, bromo, and alkoxy of from 1 to 4 carbon atoms,

an amino acid or polypeptidyl residue,

5 -OR₁₇ wherein R₁₇ is alkyl of from 1 to 4 carbon atoms, or alkyl of from 2 to 4 carbon atoms substituted with a hydroxyl group, and

 -NR₁₈R₁₉ wherein R₁₈ and R₁₉ are independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

10 each R₁₄ is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

 each R₁₅ is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

15 R₁₆ is selected from the group consisting of hydrogen, alkyl of from 1 to 4 carbon atoms optionally substituted with from 1 to 4 carbon atoms substituted with 1 or more substituents selected from the group consisting of hydroxy, chloro, bromo, and alkoxy of from 1 to 4 carbon atoms, and

 q is an integer equal to 0 or 1;

 R₃ is selected from the group consisting of hydrogen, fluoro, and hydroxy;

25 R₄ is sialyl;

 R₅ is L-fucosyl;

 Y is selected from the group consisting of O, S, -NH-, and a bond; and

 pharmaceutically acceptable salts thereof
30 and with the proviso that when R₁ is hydroxyl and R₂ is -NHC(O)CH₃ then R₃ is not hydroxyl,
 and with the further proviso that when R₁ and R₂ are hydroxyl then R₃ is not hydroxyl.

 Preferably, in Formula I, R₁ is hydroxyl, alkoxy
35 of from 1 to 4 carbon atoms, chloro, bromo, or fluoro. R₂ is preferably -NH₂, -NHC(O)R₁₃, or -N₃. R₃ is preferably hydroxyl.

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The compounds of Formula I are particularly useful in modulating a cell-mediated immune inflammatory response.

5 In another of its composition aspects, the present invention is directed to a pharmaceutical composition suitable for administration to a mammal (e.g., human) which comprises a pharmaceutically inert carrier and an effective amount of the compound of Formula I to modulate a cell-mediated immune response
10 in said mammal.

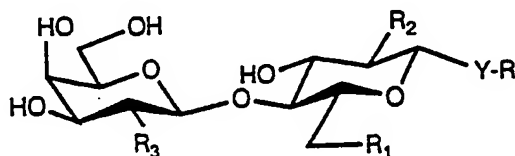
When R is an aglycon capable of being linked to a carrier, then the compounds of Formula I are also useful in preparing artificial antigens. Accordingly, in still another of its composition aspects, the
15 present invention is directed to artificial antigens prepared from a compounds of Formula I which contains an aglycon moiety capable of being linked to an antigenic carrier.

In still yet another of its composition aspects,
20 the present invention is directed to novel intermediates useful in the preparation of a compound of Formula I.

In one of its method aspects, the present invention is directed to a method for modulating a
25 cell-mediated immune response in a mammal which method comprises administering to said mammal an amount of a compound of Formula I effective in modulating said immune response.

In another of its method aspects, the present
30 invention is directed to a method for preparing a compound of Formula I which comprises

(a) selecting a compound of Formula II



II

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wherein R, R₁, R₂, R₃, and Y are as defined above;

(b) sialylating said compound at the 3 position of the galactose moiety with a β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)sialyltransferase so as to place a sialyl residue at the 3-position of the galactose unit in an α (2 \rightarrow 3)sialyl linkage; and

(c) fucosylating the compound produced in step (b) at the 3 position of the N-acetylglucosamine moiety with β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase so as to place a fucosyl residue at the 3-position of the N-acetylglucosamine unit in an α (1 \rightarrow 3) linkage.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates the structures of sialyl Lewis^x (compound 1) of analogues of sialyl Lewis^x having a substituent at the 2 position of the N-acetylglucosamine unit (compounds 12b-d) and intermediates used in preparing the analogues of sialyl Lewis^x (compounds 11b-d).

Figure 2 illustrates a general reaction scheme for the chemo-enzymatic synthesis of the analogues of sialyl Lewis^x (compound 12b-d) set forth in Figure 1 wherein Ac represent acetyl, Bn represents benzyl, and R represents $-(CH_2)_6CO_2CH_3$.

Figure 3 illustrates an alternative chemo-enzymatic synthesis of analogues of sialyl Lewis^x modified at the C-2 and/or C-6 positions of the N-acetylglucosamine unit.

Figures 4 and 5 illustrate general schemes for the synthesis of the starting materials used in the preparation of analogues of sialyl Lewis^x.

Figure 6 illustrates a general reaction scheme for the chemo-enzymatic synthesis of analogues of sialyl Lewis^x modified at the C-6 position of the N-acetylglucosamine unit.

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Figure 7 illustrates a general reaction scheme for the total chemical synthesis of analogues of sialyl Lewis^x modified at the C-2 position of the N-acetylglucosamine unit.

5 Figure 8 illustrates a general reaction scheme for preparing the 6-azido derivative of the GlcNAc saccharide unit.

10 Figure 9 illustrates a general reaction scheme for introducing the 6-alkoxy derivatives at the 6-position of GlcNAc saccharide unit.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

As noted above, the present invention is directed, in part, to the discovery of novel sialyl Lewis^x analogues which, in mammals, are useful for in vivo modulation of a cell mediated immune response. Additionally, when an appropriate aglycon is employed, these sialyl Lewis^x analogues can also be used to prepare artificial antigens for the generation of monoclonal or polyclonal antibodies to sialyl Lewis^x.

20 Additionally, the present invention is directed, in part, to novel methods for the synthesis of sialyl Lewis^x analogues.

25 However, prior to discussing this invention in further detail, the following terms will first be defined.

Definitions

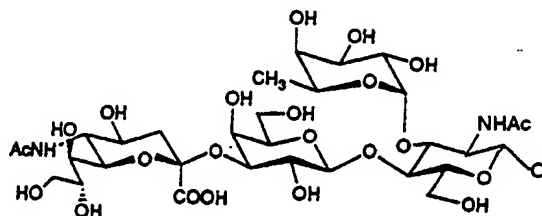
As used herein, the following terms have the definitions given below:

30 The term "cell-mediated immune response to an antigen in a mammal" refers to those mammalian immune responses which are mediated by cell-cell interactions. Included within this term are cell mediated inflammatory responses to an antigen such as

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DTH responses as well as cell-mediated inflammatory responses arising from myocardial infarction, virus-induced pneumonia, shock and sequelae (e.g., multiple organ failure), adult respiratory distress syndrome, psoriasis, arthritis, and the like. Preferably, the cell-mediated immune response is a leucocyte-mediated response.

The term "sialyl Lewis^x" (sometimes referred to "SLe^x") refers to the tetrasaccharide having the following structure:



Because of its relationship to blood group determinants, the core β Gal(1 \rightarrow 4) β GlcNAc structure of sialyl Lewis^x is often referred as a "type II structure".

The term "oligosaccharide" refers to a carbohydrate structure having from 2 to about 10 saccharide units. The particular saccharide units employed are not critical and include, by way of example, all natural and synthetic derivatives of glucose, galactose, N-acetylglucosamine, N-acetylgalactosamine, fucose, sialic acid, 3-deoxy-D,L-octulosonic acid, and the like.

In addition to being in their pyranose form, all saccharide units described herein are in their D form except for fucose which is in its L form.

The term "sialic acid" or "sialyl" means all naturally occurring structures of sialic acid and analogues of sialic acid which, as their CMP-derivatives, are compatible with the

β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)sialyltransferase. In this regard, any sialic acid which, as its CMP-derivatives, is recognized by this sialyltransferases so as to bind to the enzyme and is then available for transfer to the compound of Formula II above is said to be compatible with the sialyltransferase.

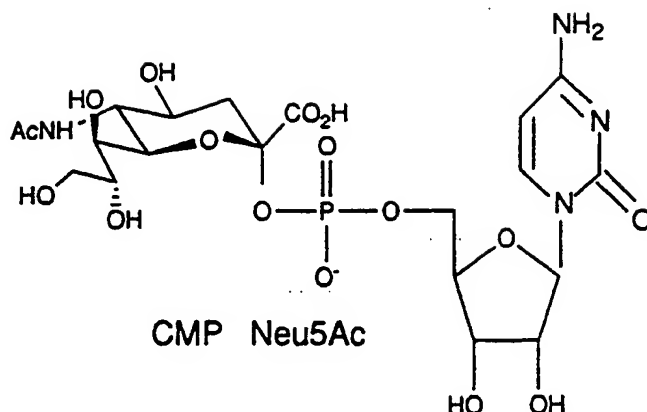
Naturally occurring structures of sialic acid include, by way of example, 5-acetamido-3,5-dideoxy-D-glycero-D-galacto-nonulopyranosylonic acid ("Neu5Ac"), N-glycolyl neuraminic acid (Neu5Gc) and 9-O-acetyl neuraminic acid (Neu5,9Ac₂). A complete list of naturally occurring sialic acids known to date are provided by Schauer³.

Analogues of sialic acid refers to analogues of naturally occurring structures of sialic acid including those wherein the sialic acid unit has been chemically modified so as to introduce and/or remove one or more functionalities from such structures. For example, such modification can result in the removal of an -OH functionality, the introduction of an amine functionality, the introduction of a halo functionality, and the like.

Certain analogues of sialic acid are known in the art and include, by way of example, 9-azido-Neu5Ac, 9-amino-Neu5Ac, 9-deoxy-Neu5Ac, 9-fluoro-Neu5Ac, 9-bromo-Neu5Ac, 7-deoxy-Neu5Ac, 7-epi-Neu5Ac, 7,8-bis-epi-Neu5Ac, 4-O-methyl-Neu5Ac, 4-N-acetyl-Neu5Ac, 4,7-di-deoxy-Neu5Ac, 4-oxo-Neu5Ac, as well as the 6-thio analogues of Neu5Ac. The nomenclature employed herein in describing analogues of sialic acid is as set forth by Reuter et al.³³

CMP-nucleotide derivative of sialic acid refers to the cytidine-5'-monophosphate derivative of a naturally occurring sialic acid or an analogue thereof. In the case where the sialic acid is Neu5Ac, the CMP derivative has the formula:

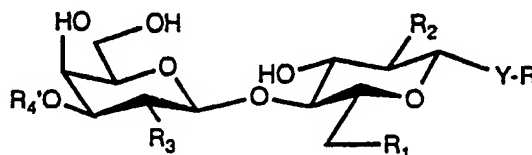
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The term "fucose" or "fucosyl" refers to L-fucose and analogues⁵⁵ thereof which, as their GDP-derivatives, are compatible with β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase. As noted below, this

5 fucosyltransferase is readily isolated from human milk²². Additionally, it is contemplated that these fucose or fucosyl compounds will be compatible with other fucosyltransferases of appropriate specificity such as cloned fucosyltransferases^{25,26}.

10 In regard to the above, any fucose compound which, as its GDP-derivative, is recognized by the β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase so as to bind to the enzyme and is then available for transfer to the compound of Formula III:



III

15 where R_4 is sialyl or hydrogen, is said to be compatible with this fucosyltransferase.

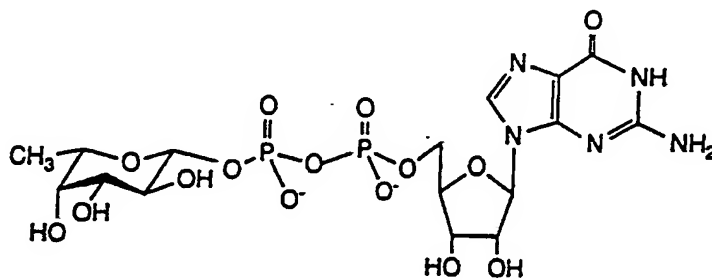
Analogues of fucose refer to naturally occurring and synthetic analogues of fucose including those

20 where the fucose unit has been chemically modified so as to introduce and/or remove one or more

functionalities from this structure. For example, such modification can result in the removal of an -OH functionality, the introduction of an amine functionality, the introduction of a halo functionality, and the like.

Certain compatible analogues of fucose are known in the art and include, by way of example, 3-deoxy-fucose, arabinose, and the like.⁵⁵

The GDP-derivative of fucose refers to guanosine 5'-(β -L-fucopyranosyl)diphosphate and any and all compatible salts thereof which has the formula:



Methods for preparing GDP-fucose are known in the art. However, GDP-fucose is preferably prepared by the method described by Jiang et al.³⁴ in U.S. Patent Application Serial No. 07/848,223 which is incorporated herein by reference in its entirety.

The term "compatible salts" as it is used in relation to guanosine 5'-(β -L-fucopyranosyl)diphosphate refers to those salts of guanosine 5'-(β -L-fucopyranosyl)diphosphate which readily form counter ions (i.e., cations) and which are compatible with the intended reactions and/or purifications. Suitable compatible salts include those prepared from counter ions such as sodium, potassium, lithium, calcium, magnesium, ammonium, mono-, di-, tri- or tetra-alkylammonium, iron, zinc, and the like.

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The term "amino acid or polypeptidyl residue" refers to product obtained by reacting an appropriate form of an amino acid or a polypeptide with an oligosaccharide glycoside of Formula I or an
5 intermediate used to prepare the oligosaccharide glycosides of Figure I and which has an amine functionality ($-NH_2$) at the 2 or 6 positions of the GlcNAc unit under conditions where the amine reacts with a carboxyl group or activated carboxyl group on
10 the amino acid or polypeptide to form an amide bond. The particular amino acid or polypeptide employed is not critical. However, in a preferred embodiment, the polypeptide contains from about 2 to about 5 amino acids and preferably from about 2 to 3 amino acids.

15 The term "pharmaceutically acceptable salts" includes the pharmaceutically acceptable addition salts of the compounds of Formula I derived from a variety of organic and inorganic counter salts well known in the art and include, by way of example only,
20 sodium, potassium, calcium, magnesium, ammonium, tetraalkylammonium, and the like.

The term "removable blocking group" refers to any group which when bound to one or more hydroxyl groups of the galactose, N-acetylglucosamine, the sialic acid
25 (including the hydroxyl group of the carboxylic acid moiety) and/or the fucose units of sialyl Lewis^x moieties prevents reactions from occurring at these hydroxyl groups and which protecting group can be removed by conventional chemical or enzymatic steps to
30 reestablish the hydroxyl group. The particular removable blocking group employed is not critical and preferred removable hydroxyl blocking groups include conventional substituents such as benzyl, acetyl, chloroacetyl, benzylidene, t-butyldiphenylsilyl and
35 any other group that can be introduced either enzymatically or chemically onto a hydroxyl functionality and later selectively removed either by

enzymatic or chemical methods in mild conditions compatible with the nature of the product. One such additional contemplated blocking group is a α -galactose which can be removed enzymatically with an α -galactosidase.

The term "aglycon of at least one carbon atom" refers to non-saccharide containing residues having at least one carbon atom. Preferably, the aglycon is selected from the group consisting of $-(A)-Z$ wherein A represents a bond, an alkylene group of from 2 to 10 carbon atoms, and a moiety of the form $-(CH_2-CR_{20}G)_n-$ wherein n is an integer equal to 1 to 5; R_{20} is selected from the group consisting of hydrogen, methyl, or ethyl; and G is selected from the group consisting of hydrogen, halogen, oxygen, sulphur, nitrogen, phenyl and phenyl substituted with 1 to 3 substituents selected from the group consisting of amine, hydroxyl, halo, alkyl of from 1 to 4 carbon atoms and alkoxy of from 1 to 4 carbon atoms; and Z is selected from the group consisting of hydrogen, methyl, phenyl, nitrophenyl and, when G is not oxygen, sulphur or nitrogen and A is not a bond, then Z is also selected from the group consisting of $-OH$, $-SH$, $-NH_2$, $-NHR_{21}$, $-N(R_{21})_2$, $-C(O)OH$, $-C(O)OR_{21}$, $-C(O)NH-NH_2$, $-C(O)NH_2$, $-C(O)NHR_{21}$, $-C(O)N(R_{21})_2$, and $-OR_{22}$ wherein each R_{21} is independently alkyl of from 1 to 4 carbon atoms and R_{22} is an alkenyl group of from 3 to 10 carbon atoms.

When the alpha-sialylated oligosaccharide glycoside is used for preparing an artificial conjugate, then the aglycon, R, is then R_{23} , which is a group capable of being linked to a carrier. Preferably, R_{23} is selected from the group consisting of $-(A)-Z'$ wherein A is selected from the group consisting of an alkylene group of from 2 to 10 carbon atoms, and a moiety of the form $-(CH_2-CR_{24}G)_n-$ wherein n is an integer equal to 1 to 5; R_{24} is selected from the

group consisting of hydrogen, methyl, or ethyl; and G is selected from the group consisting of hydrogen, oxygen, sulphur, nitrogen, phenyl and phenyl substituted with 1 to 3 substituents selected from the group consisting of amine, hydroxyl, halo, alkyl of from 1 to 4 carbon atoms and alkoxy of from 1 to 4 carbon atoms; and Z' is selected from the group consisting of hydrogen nitrophenyl and, when G is not oxygen, sulphur or nitrogen, then Z' is also selected from the group consisting of -OH, -SH, -NH₂, -NHR₂₅, -C(O)OH, -C(O)OR₂₅, -C(O)NHNH₂, and -OR₂₆ wherein each R₂₅ is independently alkyl of from 1 to 4 carbon atoms and R₂₆ is an alkenyl group of from 3 to 10 carbon atoms with the proviso that when A is a bond, Z' is not hydrogen. In such cases, the -(A)-Z' group defines a group capable of being linked to a carrier or is capable of being derivatized to a group which is capable of being linked to a carrier. The choice of an appropriate carrier may be useful in enhancing immunogenic properties.

Numerous aglycons are known in the art. For example, a linking arm comprising a para-nitrophenyl group (i.e., -YR = -OC₆H₄pNO₂) has been disclosed by Ekborg et al.⁸¹ At the appropriate time during synthesis, the nitro group is reduced to an amino group which can be protected as N-trifluoroacetamido. Prior to coupling to a support, the trifluoroacetamido group is removed thereby unmasking the amino group.

A linking arm containing sulfur is disclosed by Dahmen et al.⁸². Specifically, the linking arm is derived from a 2-bromoethyl group which, in a substitution reaction with thionucleophiles, has been shown to lead to linking arms possessing a variety of terminal functional groups such as -OCH₂CH₂SCH₂CO₂CH₃ and -OCH₂CH₂SC₆H₄-pNH₂.

Rana et al.⁸³ discloses a 6-trifluoroacetamido)-hexyl linking arm (-O-(CH₂)₆-NHCOCF₃) in which the

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trifluoroacetamido protecting group can be removed unmasking the primary amino group used for coupling.

Other exemplification of known linking arms include the 7-methoxycarbonyl-3,6,dioxaheptyl linking arm⁸⁴ $(-\text{OCH}_2-\text{CH}_2)_2\text{OCH}_2\text{CO}_2\text{CH}_3$; the 2-(4-methoxycarbonyl-butancarboxamido)ethyl⁸⁵ $(-\text{OCH}_2\text{CH}_2\text{NHC}(\text{O})(\text{CH}_2)_4\text{CO}_2\text{CH}_3)$; the allyl linking arm⁸⁶ $(\text{OCH}_2\text{CH}=\text{CH}_2)$ which, by radical co-polymerization with an appropriate monomer, leads to co-polymers; other allyl linking arms⁸⁷ are known [$-\text{O}(\text{CH}_2\text{CH}_2\text{O})_2\text{CH}_2\text{CH}=\text{CH}_2$]. Additionally, allyl linking arms can be derivatized in the presence of 2-aminoethanethiol⁸⁸ to provide for a linking arm $-\text{OCH}_2\text{CH}_2\text{CH}_2\text{SCH}_2\text{CH}_2\text{NH}_2$.

Additionally, as shown by Ratcliffe et al.²³, R group can be an additional saccharide or an oligosaccharide containing a linking arm at the reducing sugar terminus.

Preferably, the aglycon moiety is a hydrophobic group and most preferably, the aglycon moiety is a hydrophobic group selected from the group consisting of $-(\text{CH}_2)_8\text{COOCH}_3$, $-(\text{CH}_2)_5\text{OCH}_2\text{CH}=\text{CH}_2$ and $-(\text{CH}_2)_8\text{CH}_2\text{OH}$. In particular, the use of a hydrophobic group and most especially, a $-(\text{CH}_2)_8\text{COOCH}_3$, or $-(\text{CH}_2)_5\text{OCH}_2\text{CH}=\text{CH}_2$ or $-(\text{CH}_2)_8\text{CH}_2\text{OH}$ group may provide for some enhancement of the acceptor properties for transfer sialic acid by this sialyltransferase.

The carrier is a low or high molecular weight, non-immunogenic or antigenic carrier including the linking to a fluorescent label, a radioactive label, biotin, or a photolabile linking arm or a moiety to be targeted. Preferably, the carrier is an antigenic carrier and accordingly, the artificial conjugate is an artificial antigen. In some cases it may be advantageous to employ a non-immunogenic carrier.

On the other hand, the carrier can be a low molecular weight carrier such as ethylene diamine, hexamethylene diamine, tris(2-aminoethyl)amine,

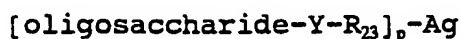
-27-

L-lysillysine, poly-L-lysine, and polymers of various molecular weights.

The term "antigenic carrier" refers to a carrier containing one or more functional groups which permit
5 linking of an oligosaccharide glycoside of Formula I (where $R = R_2$) to the carrier and which produces an antigenic response when injected into animals to which the particular carrier is not endogenous. Such carriers can be proteins [e.g., bovine serum albumin
10 (BSA), human serum albumin (HSA), diphtheria or tetanus toxoid, S-layers, and the like] and are sometimes referred to herein by the abbreviation "Ag".

The particular antigenic carrier selected for use in preparing an artificial antigen is not critical
15 provided it contains or can be derivatized to contain one or more functional groups which permit linking to the carrier of such an oligosaccharide glycoside. Suitable functional groups include, by way of example, carboxylic acid groups, amines groups (including
20 primary and secondary amines), hydroxyl groups, thio groups, and the like. Such functional groups are commonly found on antigenic carriers (e.g., proteins contain numerous such functionality) and/or can be introduced onto such carriers via art recognized
25 methods.

Coupling of one or more of the oligosaccharide glycosides of Formula I to the antigenic carrier results in a product which is described herein as an "artificial antigen" because when injected into an
30 animal, this antigen will possess one or more non-naturally occurring oligosaccharide glycoside determinants. The artificial antigens so produced are preferably represented by the formula:



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wherein oligosaccharide-Y represents the compound of formula I above (where R is R_{23}), R_{23} and Ag are as defined above, and p is an integer equal to at least 1. In this embodiment, the artificial antigen, Ag, is
5 linked to the oligosaccharide glycoside through a functional group on the antigen which couples to a complimentary functional group on the aglycon group, i.e., the R_{23} group.

The term "antibody" refers to an immunoglobulin, or derivative thereof, having an area on the surface
10 or in a cavity which specifically binds to and is thereby defined as complementary with a particular spatial and polar organization of another molecule. The antibody can be monoclonal or polyclonal and can
15 be prepared by techniques that are well known in the art, such as, for example, immunization of a host and collection of sera or hybrid cell line technology.

In this regard, the artificial antigens described above are useful in generating antibodies which
20 recognize and are complementary to the antigenic determinants, including the sialyl Lewis^x determinants, on the antigen as well as which cross-reacts with the natural substance.

The term "natural substance" refers to a
25 naturally occurring material associated with a defined disease condition (e.g., a tumor-associated carbohydrate antigen) which material contains one or more sialyl Lewis^x analogues and which material is either non-immunogenic or weakly immunogenic in the
30 diseased mammal.

B. Methodology

As indicated above, this invention is directed, in part, to novel analogues of sialyl Lewis^x which are useful in both modulating a cell-mediated immune
35 response to an antigen as well as in providing for

artificial antigens for the creation of antibodies. These novel analogues of sialyl Lewis^x can be prepared by a variety of synthetic pathways as set forth in the figures.

5 In a preferred embodiment, the analogues of sialyl Lewis^x, as set forth in Formula I, are prepared by first synthesizing the β Gal(1 \rightarrow 4) β GlcNAc-OR backbone derivatized at the 2 and/or 6 positions of the N-acetylglucosamine unit and/or at the 2 position of
10 the galactose unit. This backbone is then sequentially sialylated and fucosylated using the β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)sialyltransferase and the β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase.

 The use of such sialyltransferases and fucosyl-
15 transferase provides for the facile synthesis of analogues of sialyl Lewis^x including those having modification on either the sialyl and/or fucosyl groups. For example, use of these transferases permits the transfer of Neu5Ac or analogues of Neu5Ac
20 to the backbone structure as well as the transfer of fucose and analogues thereof to this backbone structure.

 However, there are alternative methods for preparing these compounds including methods which rely
25 on total chemical synthesis as well as on only partial enzymatic synthesis. General schemes, as set forth in Figures 2-9, for the production of analogues of sialyl Lewis^x or intermediates therefor are now discussed in more detail.

30 In those cases where the sialyl Lewis^x analogues are to be used to prepare artificial antigens, these analogues are coupled to antigenic carriers and then injected into an appropriate animal to generate antibodies.

B1. CHEMO-ENZYMATIC SYNTHESIS OF SIALYL LEWIS^x
TETRASACCHARIDES MODIFIED AT THE C-2 OR C-6
POSITIONS OF THE N-ACETYLGLUCOSAMINE

Trisaccharide 4 set forth in Figure 2 is a known
5 compound and is disclosed by Ratcliffe et al³⁵. This
compound is then derivatized by conventional steps
well known in the art to provide for a trisaccharides
11b, 11c, and 11d described in Figure 1.

Specifically, hydrogenation (H₂) of the benzyl
10 ester (-COOBn) of trisaccharide 4 at atmospheric
pressure in ethyl acetate (CH₃CO₂C₂H₅) in the presence
of 5% palladium on carbon (Pd/C), followed by de-O-
acetylation with sodium methoxide in methanol (CH₃ONa,
CH₃OH) provided trisaccharide 11b. The use of ethyl
15 acetate as solvent is recommended in the first step in
order to leave the 2-azido group untouched. Only a
very small amount of impurity is formed in this step
which can be separated by conventional separation
techniques (e.g., chromatography).

20 Alternatively, reduction of the 2-azido group of
trisaccharide 4 by hydrogen sulfide (H₂S) in a mixture
of pyridine, water and triethylamine provided the 2-
amino trisaccharide 9. Reduction of the benzyl ester
(-COOBn) followed by de-O-acetylation (as described
25 above) lead to trisaccharide 11c.

Trisaccharide 11d is prepared by first conducting
N-propionylation of trisaccharide 9 using propionic
anhydride [(CH₃CH₂CO)₂O] in methanol (CH₃OH) to provide
for trisaccharide 10. Trisaccharide 10 was
30 accompanied by a small amount of the corresponding
3-O-propionylated material which can be separated by
conventional separation techniques (e.g.,
chromatography). Removal of the acyl and benzyl
protecting groups, as indicated above, provided the
35 trisaccharide 11d.

Trisaccharide 11c can also be derivatized by
conventional methods to provide for -NHC(O)R₁₃,

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-NHSO₃H, -N=C(R₁₄)₂, -NHCH(R₁₄)₂, -N(R₁₅)₂, and an amino acid or polypeptidyl residue derivatives by conventional methods. For example, the -NH₂ group can be reacted, using conventional techniques, with:

5 a carboxylic acid, anhydride or chloride to provide for amides (e.g., as per the formation of the propionamide of trisaccharide 7d). Alternatively, the desired acid can be activated, as reported by Inazu et al.⁷² and then reacted with the amino group. The
10 carboxylic acid, anhydride, chloride, or activated acid is selected so as to provide for an R₁₃ group (i.e., as part of the -NHC(O)R₁₃ substituent) which is hydrogen or alkyl of from 1 to 4 carbon atoms optionally substituted with one or more substituents
15 (preferably 1 to 2 substituents) selected from the group consisting of hydroxy, chloro, bromo, and alkoxy of from 1 to 4 carbon atoms,

 with an appropriate form of an amino acid or polypeptide moiety activated at the acid group as
20 reported by Bodanszky et al.⁷¹;

 with an aldehyde or ketone (of from 1 to 4 carbon atoms) at controlled pH to form an imine [-N=C(R₁₄)₂] which upon reduction (e.g., with sodium cyanoborohydride) provides for an alkylamine
25 substituent [i.e., -NHCH(R₁₄)₂] as reported by Bernotas et al.⁷³,

 with a cyclic carbonate such as ethylene carbonate or propylene carbonate which ring opens upon reaction with the amine to form a carbamate group
30 having an HO-alkylene-OC(O)NH- substituent where alkylene is from 2 to 4 carbon atoms as reported by Wollenberg et al.⁷⁸, U.S. Patent No. 4,612,132,

 with a chloroformate [i.e., ClC(O)OR₁₇] in the manner disclosed by Greig et al.⁶⁹. In this case, the
35 chloroformate has an R₁₇ group which is alkyl of from 1 to 4 carbon atoms,

with $O=C(O-C_6H_4-pNO_2)_2$ which leads to an activated intermediate which is then reacted with an amine ($HNR_{18}R_{19}$) to provide for ureas $[-NHC(O)NR_{18}R_{19}]$ as described by Piekarska-Bartoszewicz et al.⁷⁰,

5 with trimethylamine, sulfur trioxide (SO_3) so as to form the $-NHSO_3H$ group as described by Petitou⁷⁶, and

with derivatized formic acid or other materials to form a formamide ($-NH-CHO$)⁷⁴ which can be further functionalized to the isocyano ($-N=C=O$) and reduced to the deoxy derivative by tributyltin hydride (Bu_3SnH)⁷⁴.

10 Trisaccharides 11b, 11c, and 11d and derivatives derived therefrom are then fucosylated by contacting the appropriate trisaccharide with $\beta Gal(1\rightarrow3/4)\beta GlcNAc \alpha(1\rightarrow3/4)$ fucosyltransferase in the presence of GDP-fucose so as to provide tetrasaccharides 12b, 12c, and 15 12d which are analogues of sialyl Lewis^x.

Figure 3 illustrates a general scheme for preparing the sialyl Lewis^x analogues of this invention from an appropriately derivatized $\beta Gal(1\rightarrow4)\beta GlcNAc$ structure by the sequential enzymatic sialylation and fucosylation of this structure. Figure 3 only 20 illustrates modification at the 2 or 6 position of the N-acetylglucosamine (GlcNAc) structure. However, it is understood that the modifications can be combined to provide for modification at both the 2 and 6 25 position of the N-acetylglucosamine. It is further understood that while Figure 3 illustrates only a 2-hydroxyl group at the 2 position of the galactose, this position may also be substituted with hydrogen or 30 fluoro. Such substituted galactose compounds are known in the art. Substitution of these galactose compounds in the reactions depicted in the figures lead to these modified galactose units in the sialyl Lewis^x analogues.

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Enzymatic Sialylation

In Figure 3, sialylation is accomplished by use of the β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)sialyltransferase [i.e., β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)ST]. The enzymatic transfer of sialic acid onto the 3-position of galactose to form α -sialyl(2 \rightarrow 3) β Gal requires the prior synthesis (i.e., activation) to its nucleotide (CMP) derivative. Activation of sialic acid is usually done by using the enzyme CMP-sialic acid synthase which is readily available and the literature provides examples of the activation of various analogues of sialic acid such as 9-substituted Neu5Ac^{36,37,38,39-41}, 7-epi-Neu5Ac⁴², 7,8-bis-epi-Neu5Ac⁴², 4-O-methyl-Neu5Ac⁴³, 4-deoxy-Neu5Ac⁴⁴, 4-acetamido-Neu5Ac⁴⁶, 7-deoxy-Neu5Ac⁴⁰, 4,7-dideoxy-Neu5Ac⁴⁰, the 6-thio derivatives of Neu5Ac⁴⁵ and Neu5OH (KDN).

The resulting CMP-sialic acid analogue (which in Figure 3 is illustrated as the CMP derivative of Neu5Ac, i.e., CMP-Neu5Ac) is then combined with the derivatized β Gal(1 \rightarrow 4) β GlcNAc-OR compound in the presence of the β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)sialyltransferase under conditions wherein sialic acid is transferred to the 3 position of the galactose to form a α Neu5Ac(2 \rightarrow 3) β Gal linkage. Suitable conditions, known in the art, include the addition of the sialyltransferase to a mixture of the derivatized β Gal(1 \rightarrow 4) β GlcNAc-OR compound and of the CMP-sialic acid in a appropriate buffer such as 0.1 M sodium cacodylate in appropriate conditions of pH and temperature such as at a pH of 6.5 to 7.5 and a temperature between 25 and 45°C, preferably 35-40°C, while incubating for 12 hours to 4 days. The resulting sialylated product can be isolated and purified using conventional methodology comprising HPLC, ion exchange-, gel-, reverse-phase- or adsorption chromatography.

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In regard to the above, it has been shown that with the $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(2\rightarrow3)\text{sialyltransferase}$, substitution of an α -fucose group at the 3 position of the GlcNAc group of the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ structure prevents the transfer of Neu5Ac to the terminal galactose. On the other hand, substitution of a saccharide unit at the 6 position of the GlcNAc group of the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ structure has been reported to permit the transfer of Neu5Ac to the terminal galactose with this sialyltransferase⁸⁹. It has now been found that in assay reactions, affinity purified $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(2\rightarrow3)\text{sialyltransferase}$ from rat liver efficiently transfers sialic acid to the 3-position of galactose (compounds 22b,c,d; 15b,g) modified at the 2 position (R_2) or the 6 position (R_1) of the GlcNAc unit of $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$. These assays are described in the examples and the results of these assay reactions are set forth in Table I below:

TABLE I
 20 Acceptor specificity of the $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(2\rightarrow3)\text{-sialyltransferase}$

	Compound			%Rate of Incorporation
	No.	R^2	R^1	
Type II	22a	NHAc	OH	100
	22b	N ₃	OH	121 ^b
	22c	NH ₂	OH	74 ^b
	22d	NHPr	OH	160 ^b
	15b	NHAc	H	47 ^b
	15g	NHAc	αFuc	106 ^c
	25 ^a measured at 2 mM acceptor concentration ^b relative to the parent type II compound (22a) β -glycoside ($R=(\text{CH}_2)_8\text{CO}_2\text{CH}_3$) ^c relative to the β -methyl glycoside of N-acetyllactosamine			

The data reported in Table I indicate that derivatives of the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc-OR}$ structure

modified at the 2 or 6 positions of the GlcNAc are active with this sialyltransferase so as to accept sialic acid at the 3 position of the galactose unit and to place a sialic acid group at an $\alpha(2\rightarrow3)$ position of the galactose. This data further shows that in some cases, the relative rate of transfer of sialic acid onto the modified $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ structure is surprisingly higher than for the unmodified structure.

Enzymatic fucosylation

In Figure 3, fucosylation is accomplished by use of the $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(1\rightarrow3/4)\text{fucosyltransferase}$ [i.e., $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(1\rightarrow3/4)\text{FT}$]. The enzymatic transfer of fucose onto the 3-position of GlcNAc to form $\alpha\text{Fuc}(1\rightarrow3)\beta\text{GlcNAc}$ requires the prior synthesis of its nucleotide (GDP) derivatives. Synthesis of GDP-fucose is preferably accomplished in the manner recited by Jiang et al.³⁴ and which is exemplified in the examples hereunder.

GDP-fucose (GDP-Fuc) is then combined with the sialylated $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc-OR}$ compound in the presence of the $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(1\rightarrow3/4)\text{fucosyltransferase}$ under conditions wherein fucose is transferred to the 3 position of the GlcNAc unit of the sialylated $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc-OR}$ compound so as to form a $\alpha\text{Neu5Ac}(2\rightarrow3)\beta\text{Gal}(1\rightarrow4)[\alpha\text{Fuc}(1\rightarrow3)]\beta\text{GlcNAc-OR}$ compound (when the sialic acid is αNeu5Ac) derivatized in the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ backbone. Suitable conditions, known in the art, include the addition of the fucosyltransferase to a mixture of the derivatized $\alpha\text{Neu5Ac}(2\rightarrow3)\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc-OR}$ compound (when the sialic acid is αNeu5Ac) and of the GDP-fucose in an appropriate buffer such as 50 mM sodium cacodylate in appropriate conditions of pH and temperature such as at a pH of 6.5 and a temperature between 30° and 45°C, preferably 35°-40°C, while

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incubating for 12 hours to 4 days. The resulting sialylated and fucosylated product can be isolated and purified using conventional methodology comprising HPLC, ion exchange-, gel-, reverse-phase- or
5 adsorption chromatography.

In the case of trisaccharides 11b-d, preparative fucosylation of these trisaccharides was performed according to Palcic et al.²² The products were purified as indicated therein. The structures of
10 trisaccharides 11b-d were confirmed by ¹H-n.m.r. at 300 MHz (Table II), and those of the resulting sialyl Lewis^x compounds, 12b-d, by ¹H-n.m.r. at 500 MHz (Tables II) below.

TABLE II

Selected ¹H-n.m.r. data for compounds: 11b, 11c, 11d, 12b, 12c, 12d^a.

Sugar Unit		Chemical Shifts (τ in Hz)					
Hydrogen		11b	11c	11d	12b	12c	12d
β-GlcR ¹	1	4.565 ^c (8.0)	4.715 (8.5)	4.552 ^c (8.0)	4.597 (8.2)	4.543 (7.8)	4.523 ^c (7.5)
	2	3.314 (8.5;10.0)	3.007 (~9.0)	--	3.45	3.065 (~9.0)	3.88
	COCH ₃	--	--	2.291 (7.5)	--	--	2.287 (7.5)
α-Fuc	COCH ₃ CH ₃	--	--	1.123	--	--	1.120
	1	--	--	--	5.398 (4.0)	5.126 (2.4)	5.100 (3.7)
	2	--	--	--	3.76	3.83	3.66
β-Gal	5	--	--	--	4.826 (6.5)	4.734 (6.5)	4.820 (6.7)
	6	--	--	--	1.165	1.195	1.166
	1	4.531 ^c (8.0)	4.542 (8.0)	4.528 ^c (8.0)	4.503 (7.6)	4.614 (8.0)	4.523 ^c (7.7)
α-Neu5Ac	2	--	--	--	3.50	3.56	3.51
	3	4.108 (3.0;10.0)	4.113 (3.0;10.0)	4.113 (3.0;10.0)	4.081 (3.0;10.0)	4.095 (3.0;10.0)	4.083 (3.0;10.0)
	4	--	3.955	3.956	3.93	--	3.92
	3 _{eq}	2.755 (4.5;12.6)	2.760 (4.5;12.6)	2.757 (4.5;12.5)	2.76 (4.4;12.5)	2.769 (4.5;12.5)	2.764 (4.5;12.5)
NHAc	3 _{ax}	1.795 (12.0)	1.795 (12.0)	1.797 (12.0)	1.795 (12.0)	1.796 (12.2)	1.793 (12.3)
	4	--	--	--	3.67	3.68	3.70
	2.029	2.031	2.031	2.029	2.030	2.025	
	CH ₃ CO ₂	2.385 (7.5)	2.387 (7.5)	2.384 (7.5)	2.387 (7.5)	2.386 (7.5)	2.384 (7.5)
CO ₂ CH ₃	3.686	3.686	3.686	3.687	3.686	3.687	

^a in D₂O, with acetone set at 2.225. ^b 11b, 11c, 11d: 300 MHz; 12b, 12c, 12d: 500 MHz. ^c interchangeable

In order to ascertain the effect on enzymatic fucosylation at the 4 position of N-acetylglucosamine arising from modification of the 2 or 6 positions of the N-acetylglucosamine of

5 α Neu5Ac(2 \rightarrow 3) β Gal(1 \rightarrow 4) β GlcNAc-OR or β Gal(1 \rightarrow 4) β GlcNAc-OR the relative rates of fucosylation of compounds 11a, 11b, 11c, and 11d as well as compound 15g were analyzed. In this regard, compound 15g was synthesized according to the procedures of Palcic et

10 al.⁶⁸

The relative rate of transfer of L-fucose to compounds 11a-d and 15g by the β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase from human milk was determined at 2 mM acceptor concentration according to

15 Palcic et al.²² These assays are described in the examples and the results of these assay reactions are summarized in Table III below:

TABLE III

20 Acceptor specificity of the fucosyltransferase isolated from human milk

	Compound			Rate of Incorporation ^a
	No.	R ²	R ⁶	%
Type II	11a	NHAc	OH	100
	11b	N ₃	OH	83
	11c	NH ₂	OH	45
	11d	NHPr	OH	105 ^b
	15g	NHAc	α Fuc(1 \rightarrow 6)	104
^a measured at 2 mM acceptor concentration				
^b relative to β Gal(1 \rightarrow 4) β GlcNAc-OR where R is $-(CH_2)_8CO_2CH_3$				

25 The above data demonstrates that modifications at the 2- and the 6-positions of the GlcNAc of the type II structures [β Gal(1 \rightarrow 4) β GlcNAc] are accepted by the

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β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase. In addition, the requirement for a free hydroxyl at the 6- position of the terminal galactose had been reported earlier⁴⁷. However, L-fucose can be

5 transferred to derivatives of type II structures where the OH at the 2, 3 and 4 of the β -galactose have been substituted by, for example, specific sugar units attached at these positions which indicates that substitution is possible at the 2-, 3- and possibly

10 the 4-hydroxyls of the terminal β -galactose when using this fucosyltransferase.

Moreover, sialylated trisaccharides modified at the C-2 position of the GlcNAc are surprisingly better acceptors than the corresponding asialo-disaccharides

15 for this fucosyltransferase.

Thus, the three-dimensional topography required by this fucosyltransferase for transfer of L-fucose to the 3 position of the GlcNAc unit of a derivatized β Gal(1 \rightarrow 4) β GlcNAc structure comprises (1) the

20 6-hydroxyl group on the galactose unit and the 3-hydroxyl group on the GlcNAc. This knowledge of the specificity of the β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4)fucosyltransferase, or of similar fucosyltransferases from other origins, for the β Gal(1 \rightarrow 4) β GlcNAc disaccharide

25 dramatically increases the usefulness of the versatile enzymes for synthetic purposes.

Consequently, the combined data from Tables I and III demonstrate that the sequential enzymatic transfer of sialic acid and L-fucose to β Gal(1 \rightarrow 4) β GlcNAc-OR

30 modified at the 2 and/or 6 positions of the GlcNAc, will proceed as in the case of the unmodified (natural) structure. Accordingly, chemical modifications at 2 and 6 positions of GlcNAc unit of β Gal(1 \rightarrow 4)GlcNAc should lead to compounds that are

35 substrates for the sialyl- and fucosyl-transferases.

As shown below, the analogues of sialyl Lewis^x modified at the 2 and/or 6 positions of the GlcNAc

unit can also be made by total chemical synthesis of these compounds.

Figure 4 illustrates the chemical synthesis of specific disaccharide derivatives of $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ starting materials starting with saccharide monomers. In this regard, the chemical coupling of the galactose and GlcNAc units results in the formation of both $\beta\text{Gal}(1\rightarrow3)\beta\text{GlcNAc}$ (type I backbone) and $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ (type II backbone) which can be separated by conventional purification techniques (i.e., chromatography).

Specifically, in Figure 4, the known³⁵ 2-azido compound 16 is protected at the 6 position with a removable protecting group (i.e., $\text{Si}(\text{C}_6\text{H}_5)_2\text{tBu}$) by conventional techniques³⁵. This derivative 17 is then combined with a fully acylated derivative of galactose 18 in the presence trimethylsilyltrifluoromethanesulfonate (TMSOTf). Afterwards, the recovered product is treated in a separate step with ammonium chloride (NH_4Cl), potassium fluoride (KF) in tetrahydrofuran. The reaction yields a mixture of $\beta\text{Gal}(1\rightarrow3)\beta\text{GlcN}_3\text{-OR}$ and $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcN}_3\text{-OR}$ derivatives, 19 and 21, which are separated by conventional methods such as chromatography.

Derivative 21 is then deprotected with a mixture of sodium methoxide in methanol ($\text{CH}_3\text{ONa}/\text{CH}_3\text{OH}$) to provide for derivative 22b which can be converted to either the amine derivative 22c or the propionate (Pr) derivative 22d following similar procedures set forth above for trisaccharides 11c and 11d.

Alternatively, derivative 21 can be tosylated by conventional techniques to provide for a tosyl group at the 6-position of the GlcN_3 derivative. The tosyl derivative can then be used to form a 6-halo substituent by a substitution reaction using the appropriate nucleophilic reagent. Derivative 21 can be converted to a 6-alkoxy substituent by alkylation

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with an alkyl halide in the presence of bis-tributyltin oxide.

Additionally, while not shown in Figure 4, the 2-deoxy ($R_2 = H$) and 2-alkoxy glucose derivatives are prepared using a synthetic scheme similar to that recited by Trumtez et al.⁷⁴ Specifically, the known 3,4,6-triacetylated 1,2-ortho ester of glucose is deacylated under conventional conditions to give the 1,2-ortho ester of glucose. This compound is then converted to the 3,4,6-tribenzyl 1,2-ortho ester of glucose using conventional techniques. The 1,2-ortho ester of the resulting compound is then opened by conventional techniques to provide a protected glycosyl donor such as the 1 α -bromo-2-acetyl-3,4,6-tribenzyl derivative of glucose. This 1 α -bromo derivative is then converted to the glycoside (-OR) by conventional techniques and the 2-acetyl group is then removed. The 2-position is now ready for formation of the blocked 2-deoxy by conventional methods such as first treating with carbon disulfide and methyl iodide in the presence of one equivalent of a base to form the -C(S)SCH₃ derivative, followed by reaction with tributyltin hydride) or for the preparation of the blocked 2-alkoxy. The blocking groups are removed by conventional techniques.

Figure 5 illustrates the synthesis of the 6-deoxy β Gal(1 \rightarrow 4)GlcNAc-OR, compound 15b, and the 6-bromo β Gal(1 \rightarrow 4)GlcNAc-OR, compound 15a. The 6-deoxy compound 15b is synthesized from a known benzylidene ring blocked saccharide (8-methoxycarbonyloctyl 2-acetamido-4,6-O-benzylidene-2-deoxy- β -D-glucopyranoside)⁵¹ which is protected at the 3-hydroxy position with a removable benzoyl blocking group (Bz) by reaction with benzoic anhydride in pyridine. Further conversion of this compound by reaction with N-bromosuccinimide and barium carbonate in carbon tetrachloride (CCl₄) at 65°C leads to the 3,4-

dibenzoyl-6-bromo-GlcNAc compound. This compound is, in turn, converted to the 6-deoxy-GlcNAc by reaction with $(C_4H_9)_3SnH$ in the presence of AIBN (azo bis-isobutyronitrile) at 70-90°C followed by treatment with methanol/sodium methoxide. The resulting 6-deoxy-GlcNAc glycoside is reacted with a known 2,3,4,6-tetraacylated derivative of galactose having an appropriate leaving group at the 1 position to permit formation of a β linkage. Suitable leaving groups include α -bromo and α -trichloroacetamidate [α -OC(=NH)CCl₃]. The reaction is conducted in the presence of a catalyst which facilitates β linkage formation. Suitable catalysts include silver trifluoromethane sulfonate in the presence of tetra-N-methyl urea when the precursor is a galactosyl bromide; and boron trifluoride etherate when the donor is galactosyl trichloroacetamidate. The reaction leads to a mixture β Gal(1 \rightarrow 3) β GlcNAc-OR and β Gal(1 \rightarrow 4)GlcNAc-OR protected compounds which can be isolated and separated by conventional techniques (e.g., chromatography). Removal of the removal protecting groups then leads to compound 15b.

As also shown in Figure 5, the debenzoylated 6-bromo-GlcNAc glycoside precursor can be reacted with a known 2,3,4,6-tetraacylated derivative of galactose having an appropriate leaving group at the 1 position to permit formation of a β linkage so as to provide for a route to the 6-bromo compounds. Suitable leaving groups include α OC(=NH)CCl₃, and the reaction is conducted in the same manner as that employed to prepare compound 15b.

Figures 6 and 7 illustrate the chemical synthesis of α -sialyl(2 \rightarrow 3) β Gal(1 \rightarrow 4) β GlcNAc-OR derivatives modified at the 6-position (Figure 6) or the 2-position (Figure 7) of the GlcNAc derivative by using one of the procedures described in Ratcliffe et al.³⁵.

Specifically, as illustrated in Figure 6, the appropriate 6-substituted derivatives of GlcNAc are prepared as above from either known⁵¹ glycoside 27 or from the known benzylidene ring blocked saccharide protected form depicted in Figure 5 (which is derived from glycoside 27) as described in detail above. The 6-derivatized blocked material (as depicted in Figure 5) is then deblocked using conventional methods to provide for compound 28 which is a 6-derivative of GlcNAc.

Compound 28 is then combined with disaccharide 29b in a manner known in the art to provide for trisaccharides 30 and 31 having conventional removable blocking groups on the Neu5Ac and the galactose units. Specifically, compound 29b is synthesized from the disaccharide 29a³⁵ by known methods and is then reacted with compound 28 in the presence of an appropriate catalyst such as $[BF_3, (C_2H_5)_2O]$ to give a mixture of the corresponding $\beta(1\rightarrow3)$ or $\beta(1\rightarrow4)$ linked trisaccharides 30 and 31, respectively. The ratio of compounds 30:31 will depend upon the nature of the substituent R_1 and on the reaction conditions. In any event, trisaccharides 30 and 31 are typically separated and purified by conventional techniques including chromatography. Removal of the blocking groups on trisaccharide 31 is also conventional (i.e., addition of hydrogen in the presence of palladium on carbon followed by treatment with sodium methoxide in the presence of methanol) and leads to the trisaccharide $\alpha\text{Neu5Ac}(2\rightarrow3)\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc}$ 25. Fucosylation of this trisaccharide 25 is preferably conducted with GDP-fucose (GDP-Fuc) in the presence of $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(1\rightarrow3/4)\text{fucosyltransferase}$ [$\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(1\rightarrow3/4)\text{FT}$] to lead to sialyl Lewis^x analogues 26 modified at the 6-position of the GlcNAc unit.

When the R_1 substituent is azido ($-N_3$ -- the synthesis of which is described below), this substituent can be further functionalized to other appropriate R_1 substituents as described above either
5 at the monosaccharide level (as shown in Figure 6) or at the trisaccharide 31 level. For example, if the R_1 group of trisaccharide 31 is an azido group, then this group can be functionalized in trisaccharide 31 to provide for the amino, amido, imino, etc. substituents
10 described above.

In any event, functionalization is generally at a point in the synthesis where the to-be formed functional group does not interfere with any of the further intended reactions. For example, if an R_1
15 functional group in monosaccharide 28 would interfere with the coupling reaction between disaccharide 29b and monosaccharide 28 then this functional group can be introduced into trisaccharide 31.

In Figure 7, the appropriate 2-substituted 6-protected derivatives of GlcNAC-OR, compound 32, are
20 prepared, for example, from the known blocked saccharide 17 depicted in Figure 4.

Compound 32 is then combined with disaccharide 29a or 29b using methods known in the art such as
25 those described by Ratcliffe et al.³⁵ to provide for trisaccharides having conventional removable blocking groups on the Neu5Ac, on the Gal, and on the 6-position of the GlcNAC units. Specifically, compound 29b is synthesized from the disaccharide 29a
30 and is then reacted with compound 32 in the presence of an appropriate catalyst such as $[BF_3, (C_2H_5)_2O]$ to give a mixture of the corresponding $\beta(1\rightarrow3)$ or $\beta(1\rightarrow4)$ linked trisaccharides, respectively. The ratio of $\beta(1\rightarrow3)$ to $\beta(1\rightarrow4)$ compounds will depend upon the nature
35 of the substituent R_2 and on the reaction conditions. In any event, these trisaccharides are typically separated and purified by conventional techniques

including chromatography. Fucosylation of this protected trisaccharide is then accomplished by reaction of the trisaccharide with an appropriate fucosyl donor such as tetra-O-benzyl-fucopyranosyl bromide as recited by Ratcliffe et al.³⁵ Removal of the blocking groups on the resulting tetrasaccharide is also conventional and leads to sialyl Lewis^x analogues modified at the 2-position of the GlcNAc unit.

Alternatively and in a preferred embodiment, fucosylation is accomplished by contacting the deprotected trisaccharide with GDP-fucose (GDP-Fuc) in the presence of β Gal(1-3/4) β GlcNAc α (1-3/4)-fucosyltransferase [β Gal(1-3/4) β GlcNAc α (1-3/4)FT] to lead to sialyl Lewis^x analogues modified at the 2-position of the GlcNAc unit.

As noted above, when the R₂ substituent is azido (-N₃), this substituent can be further functionalized to other appropriate R₂ substituents as described above either at the monosaccharide level or at the protected trisaccharide level. For example, if the R₂ group of the protected trisaccharide is an azido group, then this group can be functionalized in this trisaccharide to provide for the amino, amido, imino, etc. substituents described above. Functionalization is generally at point in the synthesis where the to-be formed functional group does not interfere with any of the further intended reactions. For example, if an R₂ functional group in monosaccharide 32 would interfere with the coupling reaction between disaccharide 29b and monosaccharide 32 then this functional group can be introduced into the protected trisaccharide.

Other derivatives at the 6-position of the GlcNAc can be prepared by art recognized methods and then these compounds can be coupled to the galactose to form β Gal(1-3) β GlcNAc-OR derivatives and β Gal(1-4) β GlcNAc-OR derivatives which can be separated

by conventional techniques (e.g., chromatography).
The β Gal(1 \rightarrow 4) β GlcNAc-OR derivatives, in turn, can be
sialylated and fucosylated as described above (per
Figure 3), to provide the sialyl Lewis^x derivatives
5 modified at the 6-position.

In regard to the above, compound 28 having a
chloro, bromo or iodo substituent at the 6 position
can be prepared by direct halogenation of the
unmodified GlcNAc-OR using the methods reported by
10 Belkhouya et al.⁷⁷

The 6-azido derivatives of GlcNAc-OR can be
prepared in the manner described in Figure 8.
Specifically, GlcNAc-OR, compound 27, is converted to
the 4,6-O-p-methoxybenzylidene blocked compound 41 by
15 reaction with $(\text{CH}_3\text{O})_2\text{CH}-\text{C}_6\text{H}_4-\text{p}-\text{OCH}_3$. This compound is
then protected at the 3-hydroxyl position by reaction
with $4-\text{CH}_3\text{O}-\text{C}_6\text{H}_4-\text{CH}_2\text{Br}$ to provide for compound 42 where
X is $4-\text{CH}_3\text{O}-\text{C}_6\text{H}_4-\text{CH}_2-$. Compound 42 is partially
deprotected at the 4 and 6 positions by reaction with
20 acetic acid (AcOH) in water at about 45°C to provide
for compound 43. The 6-mesylate, compound 44, is
prepared by reacting compound 43 with mesyl chloride
in pyridine (MsCl/py). The 6-azido derivative,
compound 45, is then formed by reaction with sodium
25 azide in dimethylformamide (DMF) and removal of the
3-blocking group with dichlorodicyanoquinone (DDQ)
yields compound 46.

The 6-mesyl compound 44 can be derivatized to any
of a number of 6-substituents including halo
30 substituents, alkoxy substituents, ester substituents,
sulfate etc. by well known chemistry.

The 6-azido compound 45 can be derivatized to
the 6-amino compound at an appropriate point in the
synthesis of the sialyl Lewis^x analogue in the manner
described above for trisaccharide 4. The 6-amino
35 derivative can then be further functionalized by
conventional methods to provide for $-\text{NHC}(\text{O})\text{R}_6$, $-\text{NHSO}_3\text{H}$,

$-N=C(R_7)_2$, $-NHCH(R_7)_2$, $-N(R_8)_2$, and an amino acid or polypeptidyl residue derivatives by conventional methods. For example, the $-NH_2$ group can be reacted, using conventional techniques, with:

5 a carboxylic acid, anhydride or chloride to provide for amides (e.g., as per the formation of the propionamide of trisaccharide 7d). Alternatively, the desired acid can be activated, as reported by Inazu et al.⁷² and then reacted with the amino group. The
10 carboxylic acid, anhydride, chloride, or activated acid is selected so as to provide for an R_6 group (i.e., as part of the $-NHC(O)R_6$ substituent) which is hydrogen or alkyl of from 1 to 4 carbon atoms optionally substituted with one or more substituents
15 (preferably 1 to 2 substituents) selected from the group consisting of hydroxy, chloro, bromo, and alkoxy of from 1 to 4 carbon atoms,

 with an appropriate form of an amino acid or polypeptide moiety activated at the acid group as
20 reported by Bodanszky et al.⁷¹;

 with an aldehyde or ketone (of from 1 to 4 carbon atoms) at controlled pH to form an imine [$-N=C(R_7)_2$] which upon reduction (e.g., with sodium cyanoborohydride) provides for an alkylamine
25 substituent [i.e., $-NHCH(R_7)_2$] as reported by Bernotas et al.⁷³,

 with a cyclic carbonate such as ethylene carbonate or propylene carbonate which ring opens upon reaction with the amine to form a carbamate group
30 having an HO-alkylene-OC(O)NH- substituent where alkylene is from 2 to 4 carbon atoms as reported by Wollenberg et al.⁷⁸, U.S. Patent No. 4,612,132,

 with a chloroformate [i.e., $ClC(O)OR_{10}$] in the manner disclosed by Greig et al.⁶⁹. In this case, the
35 chloroformate has an R_{10} group which is alkyl of from 1 to 4 carbon atoms,

with $O=C(O-C_6H_4-pNO_2)_2$ which leads to an activated intermediate which is then reacted with an amine ($HNR_{11}R_{12}$) to provide for ureas $[-NHC(O)NR_{11}R_{12}]$ as described by Piekarska-Bartoszewicz et al.⁷⁰,

5 with trimethylamine, sulfur trioxide (SO_3) at pH 9.5 so as to form the $-NHSO_3H$ group as described by Petitou⁷⁶ and

with derivatized formic acid or other materials to form a formamide ($-NH-CHO$)⁷⁴ which can be further functionalized to the isocyano ($-N=C=O$) and reduced to the deoxy derivative by tributyltin hydride (Bu_3SnH)⁷⁴.

10 The 6-alkoxy derivatives of GlcNAc can be prepared in the manner described in Figure 9. Specifically, GlcNAc-OR, compound 27, is reacted with $C_6H_5CH(OCH_3)_2$ in an acidic medium in acetonitrile to provide for the 4,6-diprotected benzylidene compound 47. In turn, compound 47 can be reacted with benzyl (Bn) bromide and sodium hydride in the presence of dimethylformamide at around $0^\circ C$ to provide for a

20 benzyl protecting group at the 3-position, i.e., compound 48. Deprotection at the 4,6 positions by contacting compound 48 with acetic acid and water at about $80^\circ-90^\circ C$ provides for compound 49. Reaction of compound 49 with bis[tributyltin] oxide and R_3Br (R_3 is

25 alkyl or substituted alkyl) provides for the 6-alkoxy compound 50. Conventional deprotection of the benzyl group with hydrogen over palladium/carbon yields compound 51.

The 6- SR_3 compounds are prepared from the 6-mesyl derivative, compound 44, by reaction with potassium thioacetate, $CH_3C(O)SK^+$, to give the thioacetate derivative at the 6-position. This derivative is then treated with mild base to produce the 6-SH derivative. The 6-SH can be reacted with an alkyl halide (e.g.,

35 CH_3Br) to provide the 6- SR_3 derivatives which, in turn, can be partially or fully oxidized to the 6-sulfone or

the 6-sulfoxide derivatives, $-S(O)R_8$ and $-S(O)_2R_8$ where R_8 is alkyl of from 1 to 4 carbon atoms.

The 6-fluoro compound is prepared from known chemistry⁷⁹ by reacting compound 49 with mesyl chloride
5 in pyridine to form the 6-mesylate which upon reaction with tetraethylammonium fluoride provides for the 6-fluoro derivative. Deprotection of the 3 benzyl group by hydrogen and palladium on carbon gave the 6-deoxy 6-fluoro derivative of compound 27.

10 The above reaction schemes depict a number of 2- or 6- substituted derivatives of GlcNAc. However, it is apparent that these modifications can be combined to provide for substituents at both the 2- and 6-positions. When disubstitution is desired,
15 the modifications are conducted at an appropriate point in the synthesis so as to be compatible with each other. That is to say that modification at the 2-position must be made with respect to the modification at the 6-position. This is within the
20 ordinary skill of the art.

Additionally, as noted above, the desired modifications to the 2 and/or 6 derivatized materials (especially of the 2-azido) are done at appropriate point in the synthetic route so as not to introduce a
25 functionality that is incompatible with subsequent reactions. However, in the case of the 6-substituted derivatives of GlcNAc, the β Gal(1 \rightarrow 4) linkage can be formed by using UDP-galactose and the commercial GlcNAc β (1 \rightarrow 4)galactosyl transferase, which is known to
30 accept modification at the 6 position. See Ichikawa et al., Anal. Biochem., 202:215-138 (1992).

B2. Coupling of Sialyl Lewis^x Analogues to Antigenic Carriers

i. Coupling of a Sialyl Lewis^x Derivative to an Artificial Carrier

5 Procedures for coupling (linking) oligosaccharide glycosides containing an aglycon having a functional group capable of linking to antigenic carrier so as to form an artificial antigen are documented in the literature^{49,50}. In general, such antigenic carriers
10 contain at least one complementary reactive functional group which will react with the functional group on the aglycon (or a derivative thereof). Care should be taken to ensure that the functional groups and the coupling procedure employed are compatible with the
15 nature of the sialyl Lewis^x analogue used and, in particular, with the functional groups present on this analogue (e.g., at the 2 and/or 6 position of the N-acetylglucosamine unit as well as the carboxylic group on the sialic acid unit). One suitable coupling
20 procedure documented in the art employs a ester functionality (COOR' where R' is a leaving group or is transformable into a leaving group such as an alkyl of from 1 to 6 carbon atoms) on the aglycon which is transformed into an acyl azide (-CON₃) following known
25 procedures. The acyl azide can then be coupled to an antigenic carrier following known procedures^{51,52}. As is recognized by the skilled artisan, the acyl azide process cannot be used for coupling when the modified sialyl Lewis^x analogue contains an amine substituent.

30 Another suitable procedure employs an aglycon moiety having a terminal ethylenic group, preferably an activated terminal ethylenic group, such as an allyloxy group -O-CH₂CH=CH₂, which can then be activated by known methods in order to effect coupling
35 to the carrier^{53,54}.

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Once the aglycon functional group of the sialyl Lewis^x analogue has been activated, the coupling reaction is generally conducted by adding a molar amount or a substantial molar excess of this oligosaccharide glycoside to a composition containing the carrier under conditions whereby the functional group(s) or activated functional groups (if activation is necessary) on the aglycon react with a complementary reactive functional groups on the carrier. The amount of the sialyl Lewis^x analogue added in conjunction with the number of reactive sites on the carrier dictates the number of such analogues attached to each carrier and this number will vary with the selected carrier. In general, sufficient sialyl Lewis^x analogues are added so as to provide at least 1 such substituent per carrier. Preferably, the number of substituents is from 1 to about 60 per each carrier and more preferably, the number of substituents is from about 1 to about 20 per each carrier.

The examples herein below present procedures for coupling a carrier having a reactive functional group to a sialyl Lewis^x analogue having a complementary reactive functional group on the aglycon moiety or a functional group on the aglycon moiety which is capable of being activated (derivatized) to a complementary reactive functional group. These examples are non-limiting.

- ii. Coupling of a β Gal(1 \rightarrow 4) β GlcNAc-OR Derivative to an Artificial Carrier followed by Sialylation and Fucosylation

As noted above, the coupling reactions useful in linking the sialyl Lewis^x analogues to the artificial antigen is limited by the fact that the coupling reaction employed must not affect the functional groups on the sialyl Lewis^x analogue in an unintended

manner (e.g., at its -COOH group on the sialic acid). To circumvent this restriction, it may be advantageous to first couple the β Gal(1 \rightarrow 4) β GlcNAc derivative to the antigenic carrier via its aglycon functionality and then, in a sequential manner, enzymatically transfer sialic acid and fucose to the β Gal(1 \rightarrow 4) β GlcNAc derivatives attached to the artificial antigen so as to provide for artificial antigens having pendent thereto one or more sialyl Lewis^x analogues.

10 In this embodiment, the coupling of the asialo oligosaccharide glycoside to the antigenic carrier is achieved in the same manner as described above. Likewise, the enzymatic transfer of sialic acid and fucose to the β Gal(1 \rightarrow 4) β GlcNAc derivatives attached to
15 the artificial antigen is also achieved in the same manner as described above.

iii. Coupling to Carriers other than Antigenic Carriers

Small molecular weight carriers could provide
20 di-, tri- or multivalent haptens with increased inhibitory potency. Appropriate sialylated Lewis^x polymeric carriers or co-polymerization of a sialyl Lewis^x monomer with an appropriate monomer could lead to non-immunogenic or biocompatible products.
25 Artificial liposomes or micelles could be used as antigens, drug carriers or multivalent inhibitors. Accordingly, in addition to coupling to antigenic carriers, the sialyl Lewis^x analogues described herein can be coupled to or incorporated with other carriers.
30 For example, if the aglycon moiety of such oligosaccharide glycosides contains a hydrophobic group, then the oligosaccharide glycosides can be incorporated into micelles and liposomes.

Liposomes and micelles containing sialyl Lewis^x
35 analogues are useful for antigens or inhibitors of cellular adhesion phenomena/targeting.

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Similarly, the carrier employed can be a solid phase particle containing one or more reactive functionalities and this can be reacted with one or more sialyl Lewis^x analogues containing a complementary reactive functional group on the aglycon which results in coupling to the solid phase particle so that the solid phase particle contains at least one conjugated sialyl Lewis^x analogue. Such coupling would proceed in a manner similar to that described above. In this embodiment, the resulting solid phase particles would be useful in isolating enzymes (not sialyltransferases) lectins or other biological receptors from an aqueous solution containing such materials. Solid phase particles containing reactive functional groups are well known in the art and include activated Sepharose, aminopropyl-silica, aminopropyl-CPG (controlled pore glass), aminoethyl cellulose, Trisacryl^R-NH, glass beads, polyacrylamide particles, and the like.

The sialyl Lewis^x analogues can also be coupled to larger molecular weight carriers of a polymeric nature which are chosen for their properties such as non-immunogenicity, bio-compatibility and the ability to incorporate numerous sialyl Lewis^x analogues per molecule of carrier.

Solid phase and polymeric carriers containing one or more sialyl Lewis^x analogues are also useful, for example, in competitive immunoassays wherein the solid phase or polymeric carriers are added to a sample suspected of containing the natural substance. Antibodies raised against the sialyl Lewis^x analogues and which cross-react with the natural substance are then added to the sample. Such antibodies are appropriately labeled so as to provide a detectable signal. The degree of binding of the labeled antibody to the solid phase or polymeric carrier depends on the amount of natural substance found in the sample.

After incubation, the solid phase or polymeric carrier is then isolated from the sample and the amount of antibody bound to the carrier is ascertained by measuring the signal level. Correlation of the measured signal to standards permits an assessment of the level of natural antigen in the sample.

Additionally, non-immunogenic conjugates would be useful as inhibitors of cellular adhesion phenomena where multivalent conjugates are contemplated to be more effective inhibitors than monovalent haptens.

C. Utility

The sialyl Lewis^x analogues described herein are useful for the treatment of diseases, including treating cell-mediated immune responses to an antigen including, by way of example, modulating cell-mediated inflammatory responses, and the like.

The sialyl Lewis^x analogues described herein are effective in suppressing cell-mediated immune responses to an antigen when administered at a dosage range of from about 0.5 mg to about 50 mg/kg of body weight, and preferably from about 0.5 to about 5 mg/kg of body weight. The specific dose employed is regulated by the particular cell-mediated immune response being treated as well as by the judgement of the attending clinician depending upon factors such as the severity of the adverse immune response, the age and general condition of the patient, and the like. The sialyl Lewis^x analogues can be administered orally, parenterally, rectally, transdermally, etc. Preferably, the sialyl Lewis^x analogues are administered parenterally.

Because suppression of cell-mediated immune responses by sialyl Lewis^x analogues requires initiation of the immune response, the sialyl Lewis^x analogues are generally administered to the patient at

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least about 0.5 hours after onset of the immune response and, preferably, at least about 1 hour after, and most preferably, at least about 5 hours after onset of the immune response.

5 In addition to modulating a cell-mediated immune response to an antigen, administration of the sialyl Lewis^x analogues also impart a degree of tolerance to additional challenges from the same antigen. In this regard, re-challenge by the same antigen weeks after
10 administration of the sialyl Lewis^x analogue results in a significantly reduced immune response (i.e., suppression of a cell-mediated immune response). Thus, administration of the sialyl Lewis^x analogue simultaneously imparts modulation (e.g., suppression)
15 of a cell-mediated immune response to an antigen and tolerance to future challenges with that antigen.

 The methods of this invention are generally achieved by use of a pharmaceutical composition comprising a pharmaceutically acceptable carrier and
20 an effective amount of a sialyl Lewis^x analogue. The pharmaceutically acceptable carrier includes, by way of example, water, buffered saline, etc. The effective amount of a sialyl Lewis^x analogue are those amounts which provide the above-noted dosage of the
25 oligosaccharide glycoside when administered to a patient. It is contemplated that suitable pharmaceutical compositions can additionally contain optional components such as an adjuvant, a preservative, etc.

30 It is also contemplated that suitable pharmaceutical compositions can include transdermal compositions or bandages which are well known in the art.

 Additionally, when attached to antigenic
35 carriers, the sialyl Lewis^x analogues described herein are useful as artificial antigens. Accordingly, such

analogues act as intermediates in the preparation of artificial antigens.

The artificial antigens containing the sialyl Lewis^x analogues described herein can be injected into mice, for example, so as to produce antibodies which cross-react with the natural substance. Such antibodies can be used in immunoassay techniques for the purpose of determining the presence and/or level of the natural substance in a sample suspected of containing the natural substance.

In addition to the above, such antibodies (particularly monoclonal antibodies) can be used in antibody therapy for a particular natural antigen (i.e., a natural substance). Specifically, artificial antigens containing one or more of the sialyl Lewis^x analogues described herein, may have one or more antigenic determinants located on the sialyl Lewis^x analogue which may be similar to an antigenic determinant in the natural antigen. When injected into mice, the artificial antigen produces antibodies which cross-react with the natural antigen. Such antibodies can then be collected and employed in antibody treatment for the natural antigen. Preferably, the antibodies are monoclonal antibodies. Methods of isolating a hybridoma line which generates monoclonal antibodies which recognize the antigenic determinant of the artificial antigen containing sialyl Lewis^x analogues and which cross-react with a similar antigenic determinant on the natural antigen are well known in the art. Optionally, such antibodies can be coupled to therapeutic agents to enhance their therapeutic effectiveness.

Likewise, utility for artificial conjugates other than artificial antigens has been set forth above.

The following examples are offered to illustrate this invention and are not to be construed in any way as limiting the scope of this invention.

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In these examples, unless otherwise defined below, the abbreviations employed have their generally accepted meaning:

	AB	=	AB pattern
5	ATP	=	Adenosine tri-phosphate
	ax	=	axial
	BSA	=	bovine serum albumin
	bt	=	broad triplet
	CDP	=	Cytidine di-Phosphate
10	d	=	doublet
	dd	=	doublet of doublets
	ddd	=	doublet of doublets of doublets
	DTH	=	delayed-type hypersensitivity
	eq	=	equatorial
15	i.r.	=	infra red
	KLH	=	Keyhole Limpet Hemocyanin
	m	=	multiplet
	q	=	quartet
	s	=	singlet
20	t	=	triplet
	t.l.c.	=	thin layer chromatography
	U	=	Units
	AG 1 × 8 (formate form) = ion exchange resin AG 1 x 8		
	(formate form) available from Bio-Rad		
25	Laboratories, Richmond, CA		
	Dowex 50W × 8 (H ⁺ form) = ion exchange resin Dowex 50W		
	x 8 (H ⁺ form) available from Dow		
	Chemical, Midland, MI		
	IRC-50 resin (H ⁺ form) = ion exchange resin IRC-50 (H ⁺		
30	form) available from Rohm & Haas,		
	Philadelphia, PA		

Commercially available components are listed by manufacturer and where appropriate, the order number. Some of the recited manufacturers are as follows:

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- Amersham = Amersham Canada Limited, Ontario, Canada
BioRad = Bio-Rad Laboratories, Richmond, CA
Iatron = Iatron Laboratories, Tokyo, Japan
Merck = E. Merck AG, Darmstadt, Germany
5 Millipore = Millipore Corp., Bedford, MA.
Pel-Freeze Biologicals = Pel-Freez, Rogers, Arkansas
Pharmacia = Pharmacia Biosystems Inc., Piscataway, NJ
Serva = Serva Feinbiochemica, Heidelberg, Germany
Sigma = Sigma Chemical Company, St. Louis, MS.
10 Waters = Waters Associates, Inc., Milford, MA.

EXPERIMENTAL

General Methods

- Pre-coated plates of silica gel (Merck, 60-F₂₅₄)
were used for analytical t.l.c. and spots were
15 detected by charring after spraying with a 5% solution
of sulfuric acid in ethanol. Silica gel 60 (Merck,
230-400 mesh) was used for column chromatography.
Iatrobeds were from Iatron (Order No. 6RS-8060).
Millex-GV filters (0.22 μ) were from Millipore. C₁₈
20 Sep-Pak cartridges and bulk C₁₈ silica gel were from
Waters Associates.

- Commercial reagents were used in chemical
reactions and solvents were purified and dried
according to usual procedures. Unless otherwise
25 noted, the reaction mixtures were processed by
dilution with dichloromethane and washing with a
dilute solution of sodium bicarbonate followed by
water. After drying over magnesium sulfate, the
solvents were removed by evaporation under vacuum with
30 a bath temperature of 35°C or lower when necessary.

¹H-n.m.r. were recorded at 300 MHz or 500 MHz with
either tetramethylsilane in CDCl₃ or acetone set at
2.225 in D₂O as internal standards, at ambient
temperature, unless otherwise noted. The chemical

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shifts and coupling constants (observed splitting) were reported as if they were first order, and only partial n.m.r. data are reported. ^{13}C -n.m.r. spectra were recorded at 75.5 MHz with tetramethylsilane in CDCl_3 or dioxane set at 67.4 in D_2O as reference.

Frozen rat livers were from Pel-Freeze Biologicals. CMP- [^{14}C]Neu5Ac was obtained from Amersham. Sepharose 6B, Dowex 1X8 were from Pharmacia, CDP and CMP-Neu5Ac were from Sigma, and ACS liquid scintillation cocktail from Amersham. GDP-fucose was obtained by chemical synthesis as described below. All other chemicals were of analytical grade and of commercial origin.

Preparative Example A -- Preparation of the
 $\beta\text{Gal}(1\rightarrow3/4)\text{-}\beta\text{GlcNAc}$
 $\alpha(2\rightarrow3)\text{sialyltransferase}$

The $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc } \alpha(2\rightarrow3)\text{sialyltransferase}$ [(EC 2.4.99.5) -- sometimes referred to as " $\alpha(2\rightarrow3)\text{ST}$ "] and the $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc } \alpha(2\rightarrow6)\text{sialyltransferase}$ [(EC 2.4.99.1) -- sometimes referred to as " $\alpha(2\rightarrow6)\text{ST}$ "] were extracted from rat liver (600 g) using Triton CF-54 (Sigma) according to Weinstein et al.⁵⁶ The enzymes from the Triton extract were partially purified and concentrated on Cibacron Blue F3GA-Sepharose by a reported modification⁵⁹ of Sticher et al.'s process.⁵⁷ The detergent extract (3L, 3.5 mg protein/ML) was loaded onto a column (8 x 20 cm) of Cibacron Blue F3GA (Serva) linked to Sepharose 6B (prepared according to Dean and Watson⁵⁸) equilibrated in 10 mM sodium cacodylate (pH 6.5), 0.15 M NaCl, 25% glycerol, 0.1% Triton CF-54 (buffer A) in two portions, with a wash step in between with buffer A. The column was washed with the same buffer until no further protein was eluted, and was then eluted with buffer A containing 2.0 M NaCl. Active fractions containing sialyltransferases were pooled, concentrated by

ultrafiltration on an Amicon PM 30 membrane and dialyzed against 200 volumes of buffer A. The $\alpha(2\rightarrow3)$ ST was separated from the $\alpha(2\rightarrow6)$ ST and purified by affinity chromatography as described by Mazid et al.⁵⁹ on a matrix (Le^c-Sephacrose) obtained by covalently linking the hapten⁵¹ $\beta\text{Gal}(1\rightarrow3)\beta\text{GlcNAcO}(\text{CH}_2)_8\text{COOH}$ to activated Sepharose as described by Matsumoto⁸⁰ using art recognized techniques involving the N-succinimidyl ester of the hapten. The sialyltransferases, partially purified by the above dye chromatography, containing ~160 mU of $\alpha(2\rightarrow3)$ ST and 2.4 U of $\alpha(2\rightarrow6)$ ST (about 860 mg protein) were diluted with an equal volume of buffer A containing 2.5 mM CDP at a flow rate of 5 mL/h. The column was washed with the equilibrating buffer to remove any loosely bound protein. Enzyme activity determination indicated that the $\alpha(2\rightarrow3)$ ST adsorbed strongly to the column during application and subsequent wash steps, while the bulk of the inert protein and the $\alpha(2\rightarrow6)$ ST eluted unretarded. The $\alpha(2\rightarrow3)$ ST was then eluted from the column with buffer A containing 0.2 M lactose. Fractions (2 mL each) containing the $\alpha(2\rightarrow3)$ ST were pooled and concentrated to a small volume (~1 mL) on an Amicon PM 30 membrane. The concentrate was dialyzed against 200 volumes of 50 mM sodium cacodylate (pH 6.5), 0.25 M NaCl, 50% glycerol, 0.1% Triton CF-54 and stored at -20°C. This preparation, 82,000-fold purified to a specific activity of 2.7 U/mg protein, was devoid of $\alpha(2\rightarrow6)$ ST activity when preparative sialylation using $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc-O}(\text{CH}_2)_8\text{COOCH}_3$ ⁵¹ (compound 22a) as the acceptor was carried out and the product analyzed by ¹H-n.m.r. spectroscopy and by t.l.c.

Enzyme Assays

Sialyltransferase activities were assayed following standard methodologies⁵⁶. The activities of the $\alpha(2\rightarrow3)$ ST and the $\alpha(2\rightarrow6)$ ST were determined in identical reaction mixtures with the exception of their acceptor substrates, which were compounds 20a and 22a (compound 20a corresponds to the β Gal(1 \rightarrow 3) β GlcNAc of compound 22), respectively. Incubation mixtures contained, in a total volume of 60 μ L, 9 nmol CMP-[¹⁴C]Neu5Ac (Amersham) (3,340 cpm/nmol), 2 mM acceptor substrate, 1 mg/mL BSA, and enzyme (0-0.2 mU) in 25 mM sodium cacodylate (pH 6.5) containing 0.5% Triton CF-54. After incubation at 37°C (10-30 min.), radioactive product was isolated by the procedure using Sep-Pak C₁₈ cartridges⁶⁰ (Waters). The cartridge was washed with water (30 mL) until background counts were obtained in the washes. Radiolabelled product was then eluted with methanol (2 X 5 mL) and quantitated in ACS scintillation cocktail (10 mL) with a Beckman LS-3801 scintillation counter. One unit of enzyme activity is defined as 1 μ mol of product formed per minute of incubation at saturating substrate concentrations. Protein concentrations were estimated using the method of Bradford⁶¹.

Sialyltransferase Kinetics

Initial transfer rates of the rat liver $\alpha(2\rightarrow3)$ ST were determined at a fixed concentration of oligo-saccharide acceptor substrate (2mM -- millimols) using reported methodologies^{56,22}. The standard incubation mixtures contained, in a total volume of 60 μ L, 560 μ M (micromols) CMP-[¹⁴C]-Neu5Ac (30,000 c.p.m.), 2 mM acceptor substrate, 1 mg/mL BSA and 360 μ U of enzyme in 25 mM sodium cacodylate buffer (pH 6.5) containing

0.5% Triton CF-54. After 30 min. of incubation at 37°C, the reaction was terminated by addition of 50 μ L of 10 mM CTP. The radiolabelled product was isolated by the Sep-Pak method⁶⁰ for acceptors with the hydrophobic 8-methoxycarboxyloctyl aglycone, or by ion-exchange chromatography on Dowex 1 X 8 (PO_4^{2-} , 100-200 mesh) for acceptors with the other aglycone as described by Paulson et al.⁶². Initial rates are expressed as a percentage with respect to the value obtained with $\beta\text{Gal}(1\rightarrow4)\beta\text{GlcNAc-OR}$ for the $\alpha(2\rightarrow3)\text{ST}$. In all cases, less than 15% of the $\text{CMP-}[^{14}\text{C}]\text{-Neu5Ac}$ was transferred to the product and the assays were performed in duplicate.

Preparative Example B -- Preparation of the
 $\beta\text{Gal}(1\rightarrow3/4)\beta\text{GlcNAc}$
 $\alpha(1\rightarrow3/4)\text{fucosyltransferase}$ from Human
Milk (EC 2.4.1.65)

The enzyme was purified from human milk obtained from Lewis^{a+b} donors, according to the methodology using affinity chromatography on GDP-hexanolamine Sepharose described by Palcic et al.²².

Fucosyltransferase Kinetics

Fucosyltransferase kinetics were performed as reported by Palcic et al.²² Incubation mixtures contained (in 50 μ L) 10 μ M $\text{GDP-}[^{14}\text{C}]\text{-fucose}$ (Amersham) (25,000 c.p.m.), 2 mM acceptor saccharide, 10 μ U fucosyltransferase, 8 mM MnCl_2 in 25 mM sodium cacodylate buffer (pH 6.5). After 20 min. of incubation at 37°C, the reaction was terminated by addition of 1.0 mL of 35 mM EDTA and the radiolabelled products were separated by the Sep-Pak method⁶⁰. Less than 20% of the $\text{GDP-}[^{14}\text{C}]\text{-fucose}$ was transferred to the product. Initial rates are expressed as a percentage with respect to the value obtained with $\alpha\text{Neu5Ac}(2\rightarrow3)\text{-}$

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β Gal(1 \rightarrow 4) β GlcNAc-OR or β Gal(1 \rightarrow 4) β GlcNAc-OR as set forth in Table III.

Example 1 -- Synthesis of the starting material/Synthesis of Acceptors: Compounds 20b, 20c, 22b, 22c, 22d (Figure 4)

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A. Preparation of 8-Methoxycarbonyl 2,3,4,6-tetra O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 3)-O-2-azido-2-deoxy- β -D-glucopyranoside (compound 19) and 8-methoxycarbonyl 2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-azido-2-deoxy- β -D-glucopyranoside (compound 21).

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A solution of trimethylsilyltrifluoromethanesulfonate (0.460 g, 1.94 mmol) in dry methylene chloride (5 mL) was syringed dropwise to a mixture of compound 17³⁵ (1.20 g, 1.94 mmol), compound 18 (0.757 g, 1.94 mmol) and molecular sieves 4A° (2 g) in dichloromethane stirred at 22°C. After 2 h, the reaction was stopped by addition of triethylamine, the mixture filtered and worked up as usual. The recovered material was chromatographed on silica gel (150 g) using a 3:1 mixture of hexanes and ethyl acetate as eluant providing a mixture of the β (1 \rightarrow 3) and the β (1 \rightarrow 4) disaccharide (1.10 g, 60%) which could not be separated at this stage.

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Tetraethylammonium chloride (0.196 g, 1.18 mmol) and anhydrous potassium fluoride (0.299 g, 5.15 mmol) were added to a solution of the mixture of the above disaccharides (0.460 g, 0.487 mmol) in dry acetonitrile (10 mL). After 24 h at 22°C, acetic acid (1-5 mL) was added and the solvents were evaporated in vacuo. The residue was dissolved in chloroform (20 mL), washed with a dilute solution of sodium bicarbonate followed by water. The recovered crude material was chromatographed on silica gel (36 g) using a 1:1 mixture of ethyl acetate and hexane as

eluant providing the (1→4) disaccharide 21 (157 mg, 46%) and the (1→3) disaccharide 19 (96 mg, 28%).

Disaccharide 21: $[\alpha]_D^{20} +6.9^\circ$ (c 1.0, CHCl_3) ^1H -n.m.r. (CDCl_3) 5.381 (d, 1H, $J_{3',4'}$ 3.5Hz, H-4'),
 5.222(dd, 1H, $J_{1',2'}$ 8.0Hz, $J_{2',3'}$ 10.0Hz, H-2'), 5.014 (dd, 1H, H-3'), 4.628(d, 1H, H-1'), 4.257[m, incl. H-1 (d, $J_{1,2}$ 8.0Hz)], 3.420-3.640[m, incl. CO_2CH_3 (s, 3.650)], 2.227(t, 2H, J 7.5Hz, CH_2CO_2), 2.150, 2.100 (two), 1.950(3s, 12H, 4 OAc), 1.600(m, 4H, methylenes),
 1.300(m, 8H, methylenes);

Disaccharide 19: $[\alpha]_D^{20} +7.8^\circ$ (c 1, CHCl_3) ^1H -n.m.r. (CDCl_3) 5.359 (d, 1H, $J_{3',4'}$ 3.2Hz), 5.225(dd, 1H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 10 Hz, H-2'), 5.013 (dd, 1H, H-3'), 4.524 (d, 1H, H-1'), 4.300 (d, $J_{1,2}$ 8.0Hz, H-1), 3.628(s, 3H, CO_2CH_3), 2.150, 2.080, 2.000, 1.920 (4s, 12H, 4 OAc), 1.600(m, 4H, methylenes), 1.300(m, 8H, methylenes).

For identification purposes both disaccharides were peracetylated in a mixture of pyridine and acetic anhydride.

Peracetylated derivative of 21: 5.314(dd, 1H, $J_{3',4'}$ 3.5, $J_{4',5'}$ <1Hz, H-4'), 5.047(dd, 1H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 10.0Hz, H-2'), 4.870-4.970[m, 2H, incl. H-3, 4.923(dd, $J_{2',3'}$ ~ $J_{3',4'}$ 10.0Hz) and H-3'(4.903, dd)], 4.420[m, 2H, incl. H-1'(d)], 4.300(d, $J_{1,2}$ 8.0Hz, H-1), 1.627[m, incl. CO_2CH_3 (s, 3.627)], 3.335(dd, 1H, H-2), 2.230(t, J 7.5Hz, CH_2CO_2) 2.080, 2.070, 2.050, 2.010, 1.980, 1.936(5s, 18H, 6 OAc), 1.570(m, 4H, methylenes), 1.210(m, 8H, methylenes).

Peracetylated derivative of compound 19:
 5.120(dd, 1H, $J_{3',4'}$ 3.5, $J_{4',5'}$ 1.0Hz, H-4'), 5.080 (dd, 1H, $J_{1',2'}$ 7.8, $J_{2',3'}$ 10.0Hz, H-2'), 4.980 (dd, 1H, H-3'), 4.875(dd, 1H, $J_{3,4}$ ~ $J_{4,5}$ ~ 10.0Hz, H-4) 4.715(d, 1H, H-1'), 4.257(d, 1H, $J_{1,2}$ 8.0Hz, H-1), 3.827(s, 3H, CO_2CH_3), 3.320(dd, 1H, $J_{2,3}$ 10.0Hz, H-2), 2.230(t, J 7.5Hz, CH_2CO_2), 2.080, 2.050, 2.020, 2.010, 1.970, (6s, 18H, 6

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OAc), 1.600(m, 4H, methylenes), 1.250(m, 8H, methylenes).

5 B. Preparation of 8-Methoxycarbonyloctyl β -D-galactopyranosyl-(1-3)-O-2-azido-2-deoxy- β -D-glucopyranoside (compound 20b)

A catalytic amount of a dilute solution of sodium methoxide in methanol was added to a solution of compound 19 (0.045 g, 0.064 mmol) in methanol (2 mL). After 5 h at 22°C, neutralization with Dowex 50W X 8 (H⁺ form) and filtration, the solvent was evaporated
10 in vacuo providing the pure 20b (30 mg, 88%); $[\alpha]_D^{20}$ -11.7° (c 0.65, H₂O) ¹H-n.m.r. (CD₃OD, DOH: 4.80): δ 4.45(d, 1H, J 7.0Hz) and 4.34(d, 1H, J 7.5Hz): H-1 and H-1', 3.61(s, CO₂CH₃), 2.27(t, 2H, J 7.5Hz, CH₂CO₂),
15 1.58(m, 4H) and 1.30 (m, 8H): methylenes.

C. Preparation of 8-Methoxycarbonyloctyl β -D-galactopyranosyl-(1-3)-O-2-amino-2-deoxy- β -D-glucopyranoside (compound 20c)

Compound 20b (0.018 g, 0.034 mmol) was
20 hydrogenated in the presence of 5% palladium on carbon (5 mg) in methanol (2 mL) at atmospheric pressure for 6 h. After filtration through Celite, the solvent was evaporated and the residue chromatographed on
Iatrobeds (2 g) using a mixture of chloroform and
25 methanol as the eluant providing the pure compound 20c; $[\alpha]_D^{20}$ -4.2° (c 0.48 H₂O); ¹H-n.m.r. (D₂O, DOH at 4.80): δ 4.56 and 4.48(2d, 1H each, J 7.5Hz): H-1 and H-1', 3.70(s, CO₂CH₃), 2.90(~t, 1H, J 9.5Hz, H-2)
2.42(t, 2H, J 7.5Hz, CH₂CO₂), 1.62(m, 4H) and 1.35 (m,
30 8H): methylenes.

D. Preparation of 8-Methoxycarbonyloctyl β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-azido-2-deoxy- β -D-glucopyranoside (compound 22b)

5 A catalytic amount of a dilute solution of sodium methoxide in methanol was added to a solution of compound 21 (0.027 g, 0.38 mmol) in methanol (2 mL). After 5 h at 22°C, neutralization with Dowex 50W X 8 (H⁺ form) and filtration, the solvent was evaporated
10 in vacuo. The residue was chromatographed on Iatrobeds using a 65:35 mixture of chloroform and methanol as eluant to give compound 22b (0.019 g, 92%); $[\alpha]_D^{20}$ -12.4° (c 0.73 CH₃OH); ¹H-n.m.r. (CD₃OD, DOH at 4.80): δ 4.32(d, 1H, J 7.5Hz) and 4.30(d, 1H, J
15 8.0Hz): H-1 and H-1', 3.60(s, CO₂CH₃), 3.13(dd, J_{1,2} 8.0 J_{2,3} 10.0Hz, H-2) 2.47(t, 2H, J 7.5Hz, CH₂CO₂), 1.56(m, 4H) and 1.29(m, 8H): methylenes.

E. Preparation of 8-Methoxycarbonyloctyl β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-amino-2-deoxy- β -D-glucopyranoside (compound 22c)

20 Compound 22b (0.016 g, 0.29 mmol) was hydrogenated in the presence of 5% palladium on carbon (10 mg) in methanol (5 mL) for 5 h at 22°C. After filtration through Celite, the solvent was evaporated
25 and the residue chromatographed on Iatrobeds (2.5 g) using a 8:2 mixture of chloroform and methanol as eluant providing the pure 22c (0.013 g, 86%). $[\alpha]_D^{20}$ 2.8° (c 0.42, H₂O).

F. Preparation of 8-Methoxycarbonyloctyl β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-deoxy-2-propionamido- β -D-glucopyranoside (compound 22d)

30 Compound 21 (0.017 g, 0.032 mmol) was hydrogenated in the presence of 5% palladium on carbon (5 mg) in methanol (8 mL) at atmospheric pressure for
35

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8 h. After filtration through Celite and evaporation of the solvent, the residue was dissolved in dry methanol (3 mL) containing some triethylamine (0.150 mL). Propionic anhydride (0.150 mL) was added and the mixture was stirred for 4 h at 22°C after which the solvents were evaporated to dryness. The residue was acetylated in a 2:1 mixture of pyridine and acetic anhydride (4 mL) at 22°C for 18 h. After addition of methanol, the mixture was worked up as usual and after evaporation of the solvents, the residue was chromatographed on silica gel using a 1:1 mixture of ethyl acetate and hexane as eluant providing the pure hexa-O-acetate of compound 22d; ¹H-n.m.r. (CDCl₃): 5.50(d, 1H, J 9.5Hz, NH), 5.32(~d, J_{3,4} 3.5Hz, H-4'), 5.07(m, 2H, H-2' and H-3), 4.93(dd, 1H, J_{2,3} 10.0Hz, H-3'), 4.25(m, 3H, incl. H-1 and H-1'), 3.63(s, CO₂CH₃), 2.257(t, 2H, J 7.5, CH₂CO₂), 2.137(dq, J 1.0 and 7.5Hz, NHCH₂), 2.11, 2.07, 2.02 (three), 1.93(4s, 12H, 6 OAc), 1.540(m, 4H) and 1.25(m, 8H): methylenes, 1.09 (t, 2H, NHCH₂CH₃).

The above disaccharide was de-O-acetylated in dry methanol (1 mL) containing a catalytic amount of a solution of sodium methoxide. After neutralization with Dowex 50W X 8 (H⁺ form) resin and filtration, evaporation of the solvents left the pure 22d; [α]_D²⁰ -18.0° (c 0.43, CH₃OH); ¹H-n.m.r. (CD₃OD, DOH at 4.80): δ 4.36(d, 1H, J 8.0Hz) and 4.33(d, 1H, J 7.5Hz): H-1 and H-1', 3.60(s, CO₂CH₃), 2.26(t, 2H, J 7.5Hz, CH₂CO₂), 2.18(q, 2H, J 7.5Hz, NHCOCH₂), 1.51(m, 4H) and 1.26(m, 8H): methylenes, 1.09(t, 3H, NHCOCH₂CH₃).

Example 2 -- Synthesis of Sialylated Trisaccharides
(Compounds 11b, 11c, and 11d)

- 5 A. Preparation of 8-methoxycarbonyloctyl (5-acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-O-(β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-azido-2-deoxy- β -D-glucopyranoside) (Trisaccharide 11b)

Trisaccharide 4 (28.8 mg, 0.024 mmol) was hydrogenated in ethyl acetate (1.5 mL) at 22°C in the presence of 5% palladium on carbon for 1 h to obtain the intermediate free acid. $[\alpha]_D^{20}$ -18.6° (c, 0.3, chloroform). This product was de-O-acetylated using a catalytic amount of sodium methoxide in methanol for 16 hours at 22°C and the recovered material was chromatographed on BioGel P2 providing trisaccharide 11b (10.4 mg, 55%), $[\alpha]_D^{20}$ -6.5° (c, 0.17, water). ¹H-n.m.r. data are reported in Table II above.

- 20 B. Preparation of 8-Methoxycarbonyloctyl (benzyl-5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-acetyl- β -D-galacto-pyranosyl)-(1 \rightarrow 4)-O-(6-O-acetyl-2-amino-2-deoxy- β -D-glucopyranoside) (trisaccharide 9)

25 Hydrogen sulfide is bubbled through a solution of trisaccharide 4 (400 mg, 0.32 mmol) in a mixture of pyridine (32 mL), water (4.8 mL) and triethylamine (1.3 mL). After 16 hours at 22°C, the mixture is evaporated to dryness and co-evaporated with toluene to give a crude trisaccharide (430 g). Some of this material (85.9 mg, 0.07 mmol) is chromatographed (10:1, toluene:ethanol) providing 9 (55 mg, 70%). $[\alpha]_D^{25}$ +25.9° (c, 0.22, chloroform); ¹H-n.m.r. (CDCl₃): δ 5.480(m, H-8", overlapping with 5.42(d, J 12.5Hz, benzylics), 5.340(dd, 1H, J_{6',7'} 2.5, J_{7',8'} 8.5Hz, H-7"), 5.052(m, incl. benzylics (d) and H-2' dd(J_{1',2'} 8.0, J_{2',3'} 10.0Hz) 5.000(dd, 1H, J_{3',4'} 3.5Hz, H-4'),

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4.490(d, 1H, J 10.0Hz, NH), 4.860(ddd, 1H, $J_{3'eq,4'} 4.5$, $J_{3'ax,4'} 12.0$, $J_{4',5'} 11.0$ Hz, H-4"), 4.640[m, 2H, incl., H-1' and H-3'), 3.660(s, 3H, OCH₃), 2.780(dd, $J_{2,3} 8.5$ Hz H-2), 2.604(dd, 1H, $J_{3'eq,3'ax} 13.0$ Hz), 2.300(t, J 7.5Hz, CH₂CO₂), 2.260, 2.170, 2.115, 2.080(three), 2.050, 1.985, 1.830(7s, 27H, 8 OAc, 1 NAc), 1.67 (t, 1H, $J_{3'eq,3'ax} 12.5$ Hz, J H-3ax), 1.600(m, 6H, methylenes), 1.240(m, 8H, methylenes).

C. Preparation of 8-Methoxycarbonyloctyl (5-acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-O-(β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-amino-2-deoxy- β -D-glucopyranoside (trisaccharide 11c)

A solution of the pure 9 (53 mg, 0.04 mmol) is hydrogenated in methanol for 1 h at 22°C in the presence of 5% palladium on carbon. Filtration of the catalyst and evaporation of the methanol provides the acid intermediate (44 mg), $[\alpha]_D +11.3^\circ$ (c, 0.22, water). This compound is de-O-acetylated using a catalytic amount of sodium methoxide in methanol for 24 h at 22°C. Evaporation of the solution obtained after neutralization with acetic acid left a material which is purified by chromatography on BioGel P2 to provide for trisaccharide 11c (29.5 mg, 99%), $[\alpha]_D -5.5^\circ$ (c, 0.22, water). ¹H-n.m.r. data are reported in Table II.

D. Preparation of 8-Methoxycarbonyloctyl (benzyl-5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(6-O-acetyl-2-deoxy-2-N-propionamido- β -D-glucopyranoside) (trisaccharide 10)

The crude amino compound 9 (98 mg, 0.08 mmol) is N-propionylated by adding propionic anhydride dropwise over 10 min to a solution of the crude amino trisaccharide 9 in a mixture of pyridine and water (about

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a 10:1 ratio of pyridine to water). The mixture is stirred overnight at 22°C, evaporated in vacuo and co-evaporated with toluene leaving a residue which is chromatographed (100:10, toluene:ethanol) providing

5 trisaccharide 10 (75.4 mg, 71%). $[\alpha]_D^{+10.3^\circ}$ (c, 0.17, chloroform); ^1H -n.m.r. (CDCl_3): δ 7.400 (m, 5H, aromatics), 5.543(d, 1H, J 7.5Hz, NH), 5.480(m, 1H, H-8"), overlapping with 5.440(d, 1H, J 12.5Hz, benzylic), 5.341 (dd, 1H, $J_{6,7}$ 2.5, $J_{7,8}$ 8.5Hz,

10 H-7"), 4.490-5.000[m, 3H, incl. benzylics (5.051, d, J 12.5 Hz), H-2' (5.038, dd, $J_{1,2}$ 8.09 $J_{2,3}$ 10.0Hz), and NH(d, J 10.5Hz)], 4.859(ddd, 1H, $J_{3,eq,4}$ 4.6, $J_{3,ax,4}$ 12.5, $J_{4,5}$ 10.5Hz, H-4"), 4.610-4.69[m, 2H, incl., H-1'(d) and H-3'(dd)], 3.58-3.70[m, 2H, incl., OCH_3 (s, 3.668)],

15 2.602(dd, 1H, $J_{3,eq,3'}$ 12.5Hz, H-3"eq), 2.150-2.330 [m, 10H, incl., CH_2CO_2 (t, J 7.5Hz), NHCOCH_2 (q, J 7.5Hz) and acetyls (2.260, 2.180, 2s)], 2.088(four), 2.068, 1.987, 1.838(4s, 21H, acetyls), 1.662(t, 1H, H-3"ax), 1.570(m, 6H,

20 methylenes), 1.240(m, 8H, methylenes), 1.130(t, 3H, CH_2CH_3).

E. Preparation of 8-methoxycarbonyloctyl (5-acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-O-(β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-deoxy-2-N-propionamido- β -D-glucopyranoside. (11d)

25

Trisaccharide 10 (71 mg, 0.055 mmol) is hydrogenated in the same manner as indicated in the synthesis of trisaccharide 11c to obtain the

30 intermediate product (64 mg, 97%), $[\alpha]_D^{-22.6^\circ}$ (c, 0.23, chloroform). This material was de-O-acetylated as usual and the recovered material chromatographed on BioGel P2 giving 11d (39 mg, 83%), $[\alpha]_D^{-8.5^\circ}$ (c, 0.2, water). ^1H -n.m.r. data are reported in Table II below.

Example 3 -- Preparative Fucosylation

i. Synthesis of GDP-Fucose.

A. Preparation of Bis(tetra-n-butylammonium)
hydrogen phosphate

5 Tetra-n-butylammonium hydroxide (40% aq. w/w, about 150g) was added dropwise to a solution of phosphoric acid (85% aq, w/w, 18g, 0.155 mmol) in water (150 mL) until the pH reached 7. Water was then evaporated in vacuo to give a syrup which was co-
10 evaporated with dry aceto-nitrile (2 x 400 mL) followed by dry toluene (2 x 400 mL). The resulting white solid (75g) was dried in vacuo and stored over phosphorus pentoxide under vacuum until used.

B. Preparation of β -L-Fucopyranosyl-1-phosphate

15 A solution of bis(tetra-n-butylammonium) hydrogen phosphate (58g, 127.8 mmol) in dry acetonitrile (300 mL) was stirred at room temperature under nitrogen in the presence of molecular sieves (4Å, 20g) for about one hour. A solution of tri-O-acetyl fucosyl-1-
20 bromide (freshly prepared from 31g, 93 mmol of L-fucose tetraacetate in the manner of Nunez et al.⁶⁵) in dry toluene (100 mL) was added dropwise in about 0.5 hour to the above solution, cooled at 0°C. After one more hour at 0°C, the mixture was brought to room
25 temperature and stirred for 3 hour. Tlc (1:1 toluene:ethyl acetate) indicated a main spot on the base line and several faster moving smaller spots.

The mixture was filtered over a pad of Celite (which was further washed with acetonitrile) and the
30 solvents evaporated in vacuo to give a red syrup. This material was dissolved in water (400 mL) and extracted with ethyl acetate (250 mL, twice). The aqueous layer was then evaporated in vacuo leaving a

yellowish syrup to which a solution of ammonium hydroxide (25% aq., 200 mL) was added. The mixture was stirred at room temperature for 3 hours after which tlc (65:35:8 chloroform:methanol:water) indicated a baseline spot. The solvent was evaporated in vacuo to give a yellowish syrup which was diluted with water (400 mL). The pH of this solution was checked and brought to 7, if necessary, by addition of a small amount of hydrochloric acid. The solution was slowly absorbed onto a column of ion exchange resin Dowex 2 X 8 [200-400 mesh, 5 x 45 cm, bicarbonate form which had been prepared by sequential washing of the resin with methanol (800 mL), water (1200 mL), ammonium bicarbonate (1 M, 1600 mL) and water (1200 mL)]. Water (1000 mL) was then run through the column followed by a solution of ammonium bicarbonate (0.5 M, 2.3 mL/minute, overnight). The eluate was collected in fractions (15 mL) and the product detected by charring after spotting on a tlc plate. Fractions 20 to 57 were pooled and evaporated in vacuo leaving a white solid which was further co-evaporated with water (3 x 300 mL) and freeze drying of the last 50 mL and then drying of the residue with a vacuum pump to give β -L-fucopyransyl-1-phosphate (9.5g, 40%) as a 12:1 mixture of β and α anomers containing some ammonium acetate identified by a singlet at $\delta=1.940$ in the ^1H -n.m.r. spectrum. This product was slowly run through a column of Dowex 5 X 8 resin (100-200 mesh, triethylammonium form) and eluted with water to provide the bis triethylammonium salt of β -L-fucopyransyl-1-phosphate as a sticky gum after freeze drying of the eluate. ^1H -n.m.r. δ : 4.840 (dd, $J_{1,2} = J_{1,P} = 7.5$ Hz, H-1), 3.82 (q, 1H, $J_{5,6} 6.5$ Hz, H-5), 3.750 (dd, 1H, $J_{3,4} 3.5$, $J_{4,5} 1.0$ Hz, H-4), 3.679 (dd, 1H, $J_{2,3} 10.0$ Hz, H-3), 3.520 (dd, 1H, H-2), 1.940 (s, acetate), 1.26 (d, H-6). Integral of the signals at 3.20 (q, $J 7.4$ Hz, NCH_2) and 1.280 and 1.260 (NCH_2CH_2)

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and H-6) indicates that the product is the bis-triethyl-ammonium salt which may loose some triethylamine upon extensive drying. ^{13}C -n.m.r. δ : 98.3 (d, $J_{\text{C,1P}}$ 3.4 Hz, C-1), 72.8 (d, $J_{\text{C,2P}}$ 7.5 Hz, C-2),
5 16.4 (C-6); ^{31}P -nmr δ : +2.6(s).

β -L-fucopyransyl-1-phosphate appears to slowly degrade upon prolonged storage (1+ days) in water at 22°C and, accordingly, the material should not be left, handled or stored as an aqueous solution at 22°C
10 or higher temperatures. In the present case, this material was kept at -18°C and dried in vacuo over phosphorus pentoxide prior to being used in the next step.

15 C. Preparation of Guanosine 5'-(β -1-fucopyranosyl)-diphosphate

Guanosine 5'-(β -1-fucopyranosyl)-diphosphate was prepared from β -L-fucopyranosyl-1-phosphate using two different art recognized procedures as set forth below:

20 PROCEDURE #1

β -L-fucopyranosyl-1-phosphate and guanosine 5'-mono-phosphomorpholidate (4-morpholine-N,N'-di-cyclohexyl-carboxamidinium salt, available from Sigma, St. Louis, Missouri, "GMP-morpholidate") were reacted
25 as described in a recent modification^{64,66} of Nunez's original procedure⁶⁵. Accordingly, tri-n-octylamine (0.800g, available from Aldrich Chemical Company, Milwaukee, Wisconsin) was added to a mixture of β -L-fucopyranosyl-1-phosphate (triethylammonium salt,
30 1.00g, about 2.20 mmol) in dry pyridine (10 mL) under nitrogen the solvent removed in vacuo. The process was repeated three times with care to allow only dry air to enter the flask. GMP morpholidate (2.4g, about 3.30 mmol) was dissolved in a 1:1 mixture of dry

dimethylformamide and pyridine (10 mL). The solvents were evaporated in vacuo and the procedure repeated three times as above. The residue was dissolved in the same mixture of solvents (20 mL) and the solution
5 added to the reaction flask accompanied by crushed molecular sieves (2g, 4Å). The mixture was stirred at room temperature under nitrogen. Tlc (3:5:2 25% aq. ammonium hydroxide, isopropanol and water) showed spots corresponding to the starting GMP-morpholidate
10 (Rf-0.8, U.V.), guanosine 5'-(β -1-fucopyranosyl)-diphosphate (Rf-0.5, U.V. and charring), followed by the tailing spot of the starting fucose-1-phosphate (Rf-0.44, charring). Additional U.V. active minor spots were also present. After stirring for 4 days at
15 room temperature, the yellowish mixture was co-evaporated in vacuo with toluene and the yellowish residue further dried overnight at the vacuum pump leaving a thick residue (2.43g). Water (10 mL) was then added into the flask to give a yellow cloudy
20 solution which was added on top of a column of AG 50W-X12 (from Biorad) resin (100-200 mesh, 25 x 1.5 cm, Na⁺ form). The product eluted with water after the void volume. The fractions which were active, both by U.V. and charring after spotting on a tlc plate, were
25 recovered and the solution freeze-dried overnight in vacuo providing a crude material (1.96g).

This residue was dissolved in water (10 mL overall) and slowly absorbed onto a column of hydrophobic C₁₈ silica gel (Waters, 2.5 x 30 cm) which
30 had been conditioned by washing with water, methanol and water (250 mL each). Water was then run through the column (0.4 mL/min) and the eluate collected in fractions (0.8 mL) which were checked by tlc (3:5:2 25% aq. ammonium hydroxide, isopropanol and water).
35 β -L-fucopyranosyl-1-phosphate, (Rf-0.54, charring) was eluted in fractions 29 to 45. A product showing a strongly U.V. active spot (Rf-0.51) eluted mainly in

fractions 46 to 65. Other minor U.V. active spots of higher or lower Rf were observed. Fractions 59 to 86, which contained guanosine 5'-(β -1-fucopyranosyl)-diphosphate (Rf~0.62), also showed a narrow U.V. active spot (Rf~0.57). Fractions 59 to 86 were pooled and freeze-dried overnight providing 0.353g of material enriched in guanosine 5'-(β -1-fucopyranosyl)-diphosphate. ¹H-n.m.r. indicated that this material was contaminated by a small amount of impurities giving signals at δ = 4.12 and δ = 5.05.

Fractions 29 to 45 and 47 to 57 were separately pooled and freeze-dried providing recovered β -L-fucopyranosyl-1-phosphate (0.264g and 0.223g, respectively, in which the second fraction contains some impurities). Occasionally, pooling of appropriate fractions provided some amount of guanosine 5'-(β -1-fucopyranosyl)-diphosphate in good purity (¹H-n.m.r.). Generally, all the material enriched in guanosine 5'-(β -1-fucopyranosyl)-diphosphate was dissolved in a minimum amount of water and run on the same column which had been regenerated by washing with large amounts of methanol followed by water. The fractions containing the purified guanosine 5'-(β -1-fucopyranosyl)-diphosphate (tlc) were pooled and freeze dried in vacuo leaving a white fluffy material (187 mg, 16%). ¹H-n.m.r. was identical to the previously reported data⁵⁵.

PROCEDURE #2

β -L-fucopyranosyl-1-phosphate and guanosine 5'-monophosphomorpholidate (4-morpholine-N,N'-di-cyclohexyl-carboxamidate salt -- "GMP-morpholidate") were reacted in dry pyridine as indicated in the original procedure⁶⁵. Accordingly, the β -L-fucopyranosyl-1-phosphate (triethylammonium salt, 0.528g, about 1.18 mmol) was dissolved in dry pyridine

(20 mL) and the solvent removed in vacuo. The process was repeated three times with care to allow only dry air to enter the flask. GMP-morpholidate (1.2g, 1.65 mmol) and pyridine (20 mL) were added into the
5 reaction flask, the solvent evaporated in vacuo and the process repeated three times as above. Pyridine (20 mL) was added to the final residue and the heterogeneous mixture was stirred for 3 to 4 days at room temperature under nitrogen. An insoluble mass
10 was formed which had to be occasionally broken down by sonication.

The reaction was followed by tlc and worked up as indicated in the first procedure to provide the GDP-fucose (120 mg, 16%).

15 ii. Enzymatic Conditions

β Gal(1 \rightarrow 3/4) β GlcNAc α (1 \rightarrow 3/4) fucosyltransferase was purified from human milk according to the methodology using affinity chromatography on GDP-hexanolamine Sepharose described by Palcic et al.²²
20 The enzymatic reactions were carried out at 37°C in a plastic tube using a sodium cacodylate buffer (100 mM, pH 6.5), MnCl₂ (10 mM), ATP (1.6 mM) NaN₃ (1.6 mM). The final reaction mixture was diluted with H₂O (5 mL) and applied onto C₁₈ Sep-Pak cartridges as reported²².
25 After washing with H₂O (30 mL) the products were eluted with CH₃OH and the solvents evaporated. The residue was dissolved in a 65:35:5 mixture of CHCl₃, CH₃OH, and H₂O and applied on a small column of Iatrobeads (0.200 to 0.500 g). After washing with the same solvent
30 mixture, the products were eluted with a 65:35:8 and/or 60:40:10 mixtures of the same solvents. The appropriate fractions (t.l.c.) were pooled, the solvents evaporated in vacuo, the residue run through a small column of AG 50W X 8 (Na⁺ form) (BioRad) in H₂O
35 and the products recovered after freeze drying in

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vacuo. ¹H-n.m.r. data of the tetrasaccharides are reported in Table II above.

iii. Fucosylation Reactions

- 5 A. Preparation of 8-Methoxycarbonyloctyl(5-acetamido-3,5-di-deoxy-D glycer- α -D-galacto-2-nonulopyranosylonic acid)-(2-3)-O-[β -D-galactopyranosyl]-(1-4)-O-[α -L-fucopyranosyl-(1-3)-O]-(2-azido-2-deoxy- β -D-glucopyranoside) (tetrasaccharide 12b)
- 10 Trisaccharide 11b (7.7 mg), GDP-fucose (18 mg), the fucosyltransferase (20 mU) and calf intestine alkaline phosphatase (10 U) were incubated for 72 h in the buffer (2 mL). Isolation and purification provided 12b (2.84 mg).
- 15 B. Preparation of 8-Methoxycarbonyloctyl(5-acetamido-3,5-di-deoxy-D-glycer- α -D-galacto-2-nonulopyranosylonic acid)-(2-3)-O-[β -D-galactopyranosyl]-(1-4)-O-[α -L-fucopyranosyl-(1-3)-O]-(2-amino-2-deoxy- β -D-glucopyranoside) (tetrasaccharide 12c)
- 20
- Trisaccharide 11c (8.4 mg), GDP-fucose (18 mg), the fucosyltransferase (20 mU) and calf intestine alkaline phosphatase (10 U) were incubated for 67 h in the buffer (2 mL). Isolation and purification
- 25 provided 12c (2.46 mg).
- C. Preparation of 8-Methoxycarbonyloctyl(5-acetamido-3,5-di-deoxy-D-glycer- α -D-galacto-2-nonulopyranosylonic acid)-(2-3)-O-[β -D-galactopyranosyl]-(1-4)-O-[α -L-fucopyranosyl-(1-3)-O]-(2-N-propionamido-2-deoxy- β -D-glucopyranoside) (tetrasaccharide 12d)
- 30
- Trisaccharide 11d (8.3 mg), GDP-fucose (18 mg), the fucosyltransferase (18 mU) and calf intestine
- 35 alkaline phosphatase (10 U) were incubated for 72 h in

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the buffer (2 mL). Isolation and purification provided **12d** (6.17 mg).

Example 5 -- SYNTHESIS OF ARTIFICIAL ANTIGENS

5 Sialyl Lewis^x analogues containing an appropriate functional group in the glycoside moiety can be conjugated to antigenic carriers containing one or more complementary functional groups following procedures known in the art. Depending on the linking
10 chemistry employed, the sialyl and fucosyl groups can be included in the saccharide to be conjugated or, a β Gal(1 \rightarrow 4) β GlcNAc derivative can be conjugated and the resulting conjugate sialylated and fucosylated to provide for sialyl Lewis^x analogues.

15 Specifically, the conjugation of the disaccharide glycosides such as structure **22b** to BSA, KLH or other carriers is achieved by procedures such as the "acyl-azide" procedure of Lemieux et al. and Pinto et al.^{51,52}. The resulting conjugates are then sialylated and
20 fucosylated in a manner similar to that reported above and the products purified by a combination of ultra-filtration and Gel-filtration to provide for artificial antigens having one or more sialyl Lewis^x analogues attached thereto.

25 Similarly, other antigenic carriers can be used to create artificial antigens including KLH, human serum albumin (HSA), diphtheria or tetanus toxins, S-layers, and the like. Likewise, other derivatives of β Gal(1 \rightarrow 4) β GlcNAc could be conjugated (coupled) to
30 the antigenic carriers used in place of compound **22b**.

 Alternatively, tetrasaccharide **12b** can be directly conjugated to antigenic carriers containing one or more complementary functional groups following procedures known in the art. For example, the
35 $-(CH_2)_nCO_2CH_3$ aglycon of tetrasaccharide **12b** can be modified by reaction with hydrazine and N_2O_4 to convert

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the ester (COOCH_3) to an acyl azide ($-\text{C}(\text{O})\text{N}_3$). The acyl azide is then displaced by reaction with an amino functionality on the antigenic carrier resulting in linking of the alpha sialylated oligosaccharide glycoside to the carrier via an amide bond.

Because the carrier can contain numerous amine groups, the carrier is capable of adding more than one sialyl Lewis^x derivatives.

Example 6 -- SYNTHESIS OF AGGREGATES CONTAINING SIALYL LEWIS^x ANALOGUES

Aggregates such as liposomes and micelles can be prepared so as to incorporate sialyl Lewis^x analogues. Specifically, incorporation of the sialyl Lewis^x analogue into such aggregates requires that the aglycon moiety be sufficiently hydrophobic to be incorporated into such aggregates. It is contemplated that such hydrophobic aglycons can include the $-(\text{CH}_2)_2\text{COOCH}_3$ which has been extended by various moieties such as naphthyl, substituted naphthyl, octyl, and the like which would improve the ability to incorporate the saccharide into the aggregate.

In such aggregates, the hydrophobic aglycon group of the sialyl Lewis^x analogue becomes partitioned in the lipid portion of the aggregate whereas the tetra-saccharide group is generally partitioned in the aqueous phase.

Methods of preparing such aggregates are well known in the art. See, for instance, U.S. Patent No. 4,522,803 which is incorporated herein by reference.

Additionally, sialyl Lewis^x analogues are disclosed in our concurrently filed application, Serial No. 07/_____, filed as Attorney Docket No. 000475-011 and entitled "Modified Sialyl Lewis^x Compounds". As noted above, that application is filed

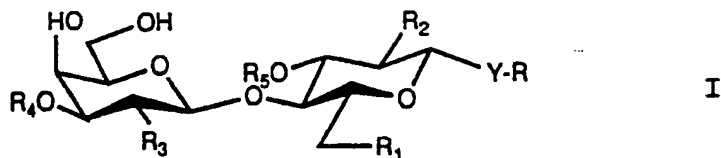
concurrently this application and is incorporated herein by reference in its entirety.

Now having fully described this invention, it will be understood by those with skill in the art
5 that the invention may be practiced within a wide and equivalent range of conditions, parameters, and the like, without affecting the scope or spirit of the invention or any embodiment thereof.

Modifications of the above-described methods for
10 carrying out the invention that are obvious to persons of skill in medicine, chemistry, biochemistry, immunology, and/or related fields are intended to be within the scope of the following claims.

WHAT IS CLAIMED IS:

1. A compound represented by Formula I:



wherein R is selected from the group consisting
 5 of hydrogen, a saccharide, an oligosaccharide, or an
 aglycon having at least 1 carbon atom;

R_1 is selected from the group consisting of
 hydrogen, $-NH_2$, N_3 , $-NHSO_3H$, $-NR_8C(O)R_6$, $-N=C(R_7)_2$,
 $-NHCH(R_7)_2$, $-N(R_8)_2$, $-SR_8$, $-O(C(O))_pR_9$, fluoro, chloro,
 10 bromo and sulfate,

wherein R_6 is selected from the group consisting
 of hydrogen,

alkyl of from 1 to 4 carbon atoms optionally
 substituted with 1 or more substituents selected from
 15 the group consisting of hydroxy, chloro, bromo, alkoxy
 of from 1 to 4 carbon atoms, phenyl, and phenyl
 substituted with 1 to 3 substituents selected from the
 group consisting of hydroxy, alkyl of from 1 to 4
 carbon atoms, alkoxy of from 1 to 4 carbon atoms,
 20 chloro, bromo, and sulfate,

an amino acid or polypeptidyl residue,

$-OR_{10}$ wherein R_{10} is alkyl of from 1 to 4
 carbon atoms, or alkyl of from 2 to 4 carbon atoms
 substituted with a hydroxyl group, and

25 $-NR_{11}R_{12}$ wherein R_{11} and R_{12} are independently
 selected from the group consisting of hydrogen and
 alkyl of from 1 to 4 carbon atoms,

each R_7 is independently selected from the
 group consisting of hydrogen and alkyl of from 1 to 4
 30 carbon atoms,

R_8 is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

5 R_9 is selected from the group consisting of hydrogen, alkyl of from 1 to 4 carbon atoms optionally substituted with 1 or more substituents selected from the group consisting of hydroxy, chloro, bromo, alkoxy of from 1 to 4 carbon atoms, phenyl, and phenyl substituted with 1 to 3 substituents selected from
10 hydroxy, alkyl of from 1 to 4 carbon atoms, alkoxy of from 1 to 4 carbon atoms, chloro, bromo, and sulfate, and p is an integer equal to 0 or 1;

R_2 is selected from the group consisting of hydrogen, $-N_3$, $-NH_2$, $-NHSO_3H$, $-NR_{15}C(O)R_{13}$, $-N=C(R_{14})_2$,
15 $-NHCH(R_{14})_2$, $-N(R_{15})_2$, $-O(C(O))_qR_{16}$, fluoro, chloro, and sulfate,

wherein R_{13} is selected from the group consisting of hydrogen,

alkyl of from 1 to 4 carbon atoms optionally substituted with 1 or more substituents selected from
20 the group consisting of hydroxy, chloro, bromo, and alkoxy of from 1 to 4 carbon atoms,

an amino acid or polypeptidyl residue,

$-OR_{17}$ wherein R_{17} is alkyl of from 1 to 4
25 carbon atoms, or alkyl of from 2 to 4 carbon atoms substituted with a hydroxyl group, and

$-NR_{18}R_{19}$ wherein R_{18} and R_{19} are independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

30 each R_{14} is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

each R_{15} is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4
35 carbon atoms,

R_{16} is selected from the group consisting of hydrogen, alkyl of from 1 to 4 carbon atoms optionally

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substituted with from 1 to 4 carbon atoms substituted with 1 or more substituents selected from the group consisting of hydroxy, chloro, bromo, and alkoxy of from 1 to 4 carbon atoms, and

5 q is an integer equal to 0 or 1;

 R₃ is selected from the group consisting of hydrogen, fluoro, and hydroxy;

 R₄ is sialyl;

 R₅ is L-fucosyl;

10 Y is selected from the group consisting of O, S, -NH-, and a bond; and
 pharmaceutically acceptable salts thereof
 and with the proviso that when R₁ is hydroxyl
and R₂ is -NHC(O)CH₃, then R₃ is not hydroxy.

15 2. A compound of Claim 1 wherein R₃ is hydroxyl.

 3. A compound of Claim 2 wherein R₁ is hydroxyl, alkoxy of from 1 to 4 carbon atoms, fluoro, chloro, or bromo.

 4. A compound of Claim 1 wherein R₂ is selected
20 from the group consisting of -NH₂, -NHC(O)R₁₃, or -N₃.

 5. A compound of Claim 1 wherein R₄ is Neu5Ac.

 6. A compound of Claim 1 wherein R₅ is fucose.

 7. A compound of Claim 1 wherein R₁ is hydroxy,
R₂ is azido, R₃ is hydroxy, R₄ is Neu5Ac, and R₅ is
25 fucose.

 8. A compound of Claim 1 wherein R₁ is hydroxy,
R₂ is -NH₂, R₃ is hydroxy, R₄ is Neu5Ac, and R₅ is
fucose.

9. A compound of Claim 1 wherein R_1 is chloro, R_2 is $-\text{NHC}(\text{O})\text{CH}_3$, R_3 is hydroxy, R_4 is Neu5Ac, and R_5 is fucose.

10. A compound of Claim 1 wherein R_1 is bromo, R_2 is $-\text{NHC}(\text{O})\text{CH}_3$, R_3 is hydroxy, R_4 is Neu5Ac, and R_5 is fucose.

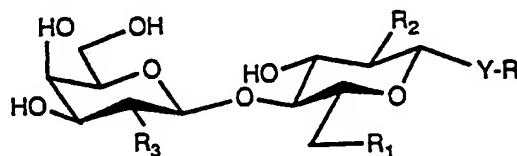
11. A compound of Claim 1 wherein R_1 is fluoro, R_2 is $-\text{NHC}(\text{O})\text{CH}_3$, R_3 is hydroxy, R_4 is Neu5Ac, and R_5 is fucose.

12. A pharmaceutical composition suitable for administration to a mammal which comprises a pharmaceutically inert carrier and an effective amount of the compound of Claim 1 to modulate a cell-mediated immune response in said mammal.

13. A method for modulating a cell-mediated immune response in a mammal which method comprises administering to said mammal an amount of a compound of Claim 1 effective in modulating said immune response.

14. A method for preparing a compound of Claim 1 which comprises

(a) selecting a compound of Formula II



II

wherein R , R_1 , R_2 , R_3 , and Y are as defined above;

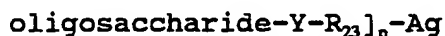
(b) sialylating said compound at the 3 position of the galactose moiety with a sialyltransferase selected from the group consisting of

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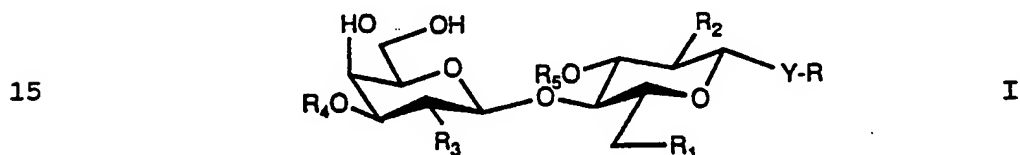
β Gal(1 \rightarrow 3/4) β GlcNAc α (2 \rightarrow 3)-sialyltransferase and
 β Gal(1 \rightarrow 3)GalNAc α (2 \rightarrow 3)sialyl-transferase so as to
 place an α (2 \rightarrow 3)sialyl residue on the galactose unit;
 and

- 5 (c) fucosylating the compound produced in step
 (b) at the 4 position of the N-acetylglucosamine
 moiety with β Gal(1 \rightarrow 3/4) β GlcNAc
 α (1 \rightarrow 3/4)fucosyltransferase so as to form an
 α (1 \rightarrow 4)fucosyl residue on the N-acetylglucosamine unit.

- 10 15. An artificial antigen represented by the
 formula:



wherein oligosaccharide is a compound represented by
 Formula I:



wherein R_1 is selected from the group consisting
 of hydrogen, $-\text{NH}_2$, N_3 , $-\text{NHSO}_3\text{H}$, $-\text{NR}_8\text{C}(\text{O})\text{R}_6$, $-\text{N}=\text{C}(\text{R}_7)_2$,
 $-\text{NHCH}(\text{R}_7)_2$, $-\text{N}(\text{R}_8)_2$, $-\text{SR}_8$, $-\text{O}(\text{C}(\text{O}))_p\text{R}_9$, fluoro, chloro,
 bromo and sulfate,

- 20 wherein R_6 is selected from the group consisting
 of hydrogen,

alkyl of from 1 to 4 carbon atoms optionally
 substituted with 1 or more substituents selected from
 the group consisting of hydroxy, chloro, bromo, alkoxy
 25 of from 1 to 4 carbon atoms, phenyl, and phenyl
 substituted with 1 to 3 substituents selected from the
 group consisting of hydroxy, alkyl of from 1 to 4
 carbon atoms, alkoxy of from 1 to 4 carbon atoms,
 chloro, bromo, and sulfate,

- 30 an amino acid or polypeptidyl residue,

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-OR₁₀ wherein R₁₀ is alkyl of from 1 to 4 carbon atoms, or alkyl of from 2 to 4 carbon atoms substituted with a hydroxyl group, and

5 -NR₁₁R₁₂ wherein R₁₁ and R₁₂ are independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

each R₇ is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

10 R₈ is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

R₉ is selected from the group consisting of hydrogen, alkyl of from 1 to 4 carbon atoms optionally substituted with 1 or more substituents selected from the group consisting of hydroxy, chloro, bromo, alkoxy of from 1 to 4 carbon atoms, phenyl, and phenyl substituted with 1 to 3 substituents selected from hydroxy, alkyl of from 1 to 4 carbon atoms, alkoxy of from 1 to 4 carbon atoms, chloro, bromo, and sulfate, and p is an integer equal to 0 or 1;

20 R₂ is selected from the group consisting of hydrogen, -N₃, -NH₂, -NHSO₃H, -NR₁₃C(O)R₁₃, -N=C(R₁₄)₂, -NHCH(R₁₄)₂, -N(R₁₅)₂, -O(C(O))_qR₁₆, fluoro, chloro, and sulfate,

25 wherein R₁₃ is selected from the group consisting of hydrogen,

alkyl of from 1 to 4 carbon atoms optionally substituted with 1 or more substituents selected from the group consisting of hydroxy, chloro, bromo, and alkoxy of from 1 to 4 carbon atoms,

30 an amino acid or polypeptidyl residue,

-OR₁₇ wherein R₁₇ is alkyl of from 1 to 4 carbon atoms, or alkyl of from 2 to 4 carbon atoms substituted with a hydroxyl group, and

35

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$-NR_{18}R_{19}$ wherein R_{18} and R_{19} are independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

5 each R_{14} is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

each R_{15} is independently selected from the group consisting of hydrogen and alkyl of from 1 to 4 carbon atoms,

10 R_{16} is selected from the group consisting of hydrogen, alkyl of from 1 to 4 carbon atoms optionally substituted with from 1 to 4 carbon atoms substituted with 1 or more substituents selected from the group consisting of hydroxy, chloro, bromo, and alkoxy of
15 from 1 to 4 carbon atoms, and

q is an integer equal to 0 or 1;

R_3 is selected from the group consisting of hydrogen, fluoro, and hydroxy;

R_4 is sialyl;

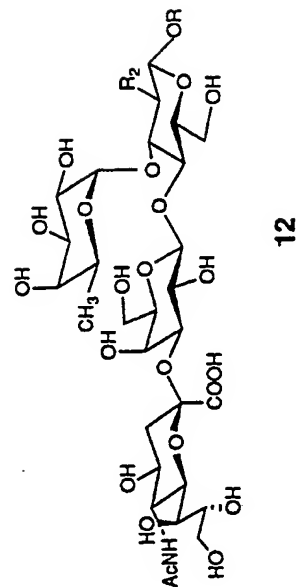
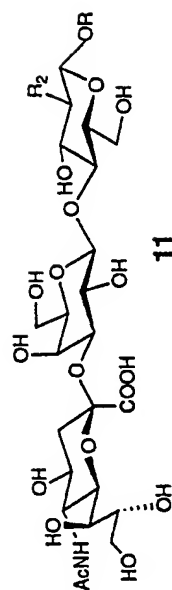
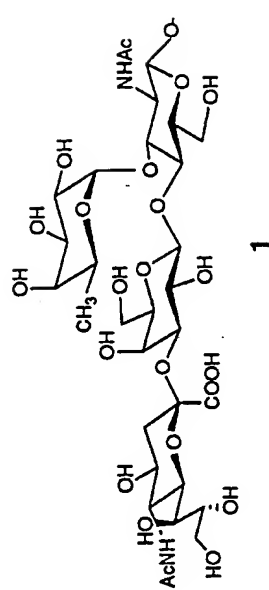
20 R_5 is L-fucosyl;

Y is selected from the group consisting of O, S, -NH-, and a bond;

R_{23} is an aglycon moiety capable of being linked to an antigenic carrier;

25 p is an integer equal to at least one; and
 Ag is an antigenic carrier.

Figure 1



R_2	NHAc	N_3	NH_2	NHPr
11a	11b	11c	11d	11d
12a	12b	12c	12d	12d

$$R = (CH_2)_8CO_2CH_3$$

$$Pr = COCH_2CH_3$$

Figure 2

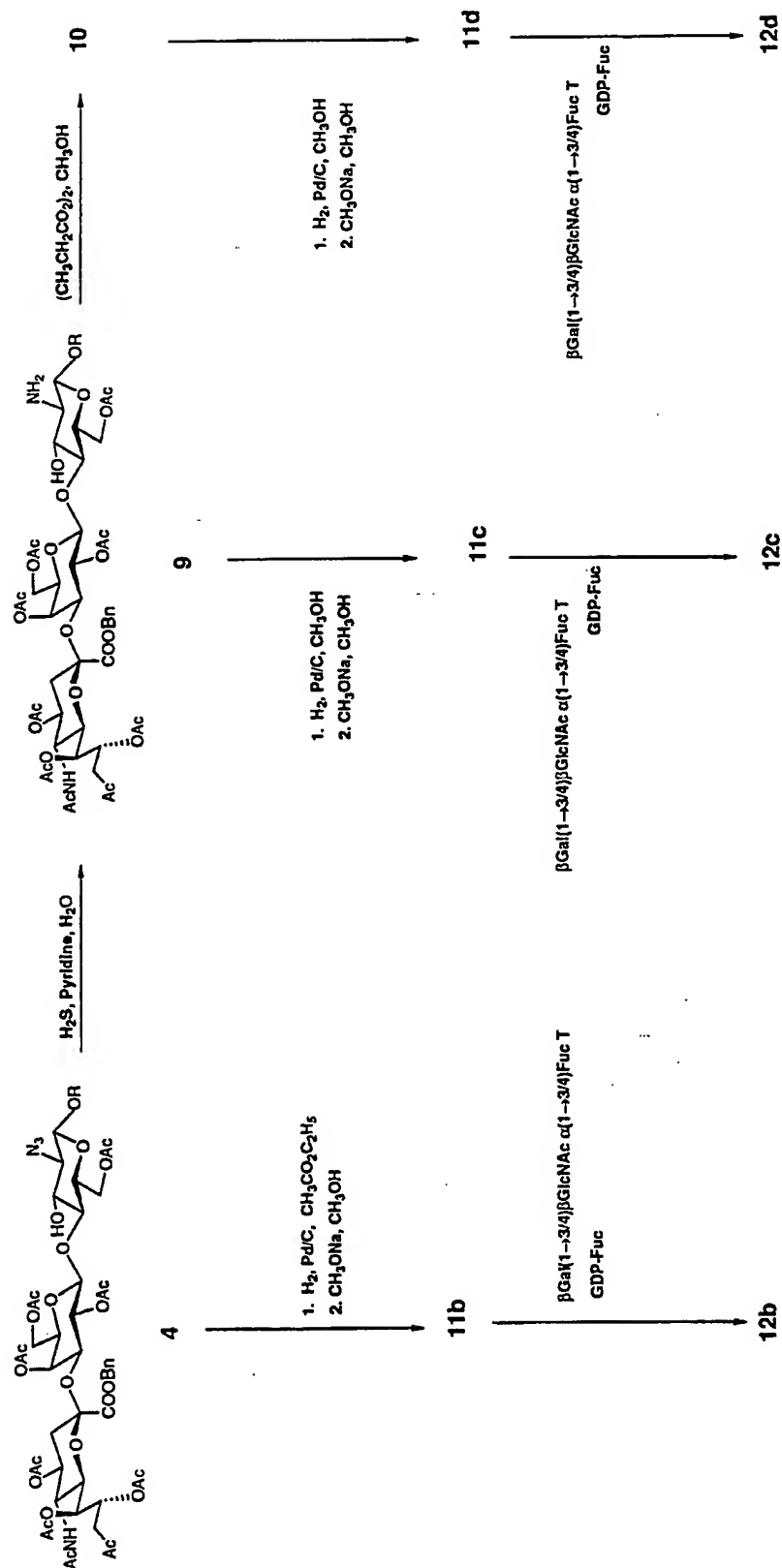
R = (CH₂)₈CO₂CH₃

Figure 3

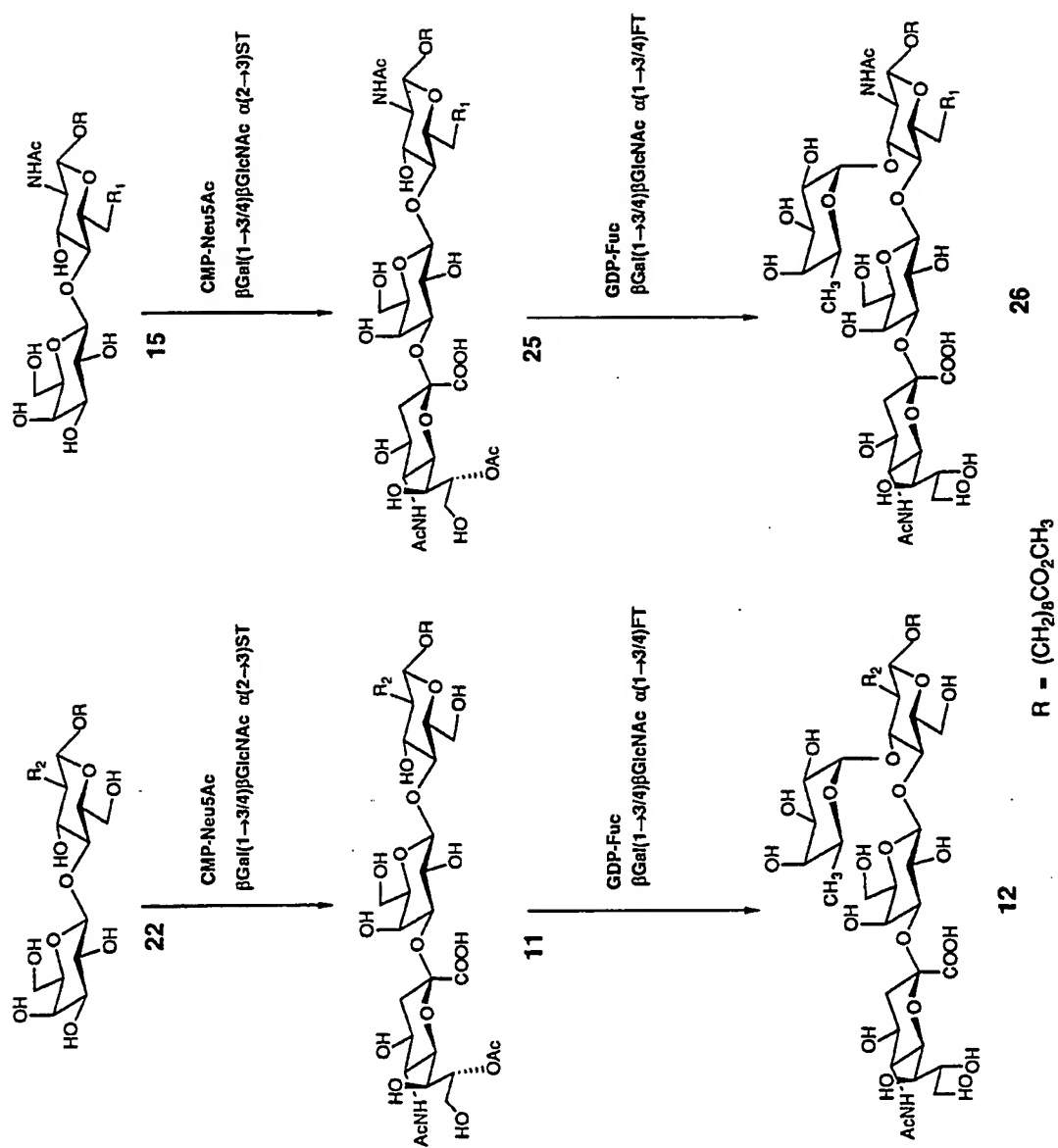


Figure 4

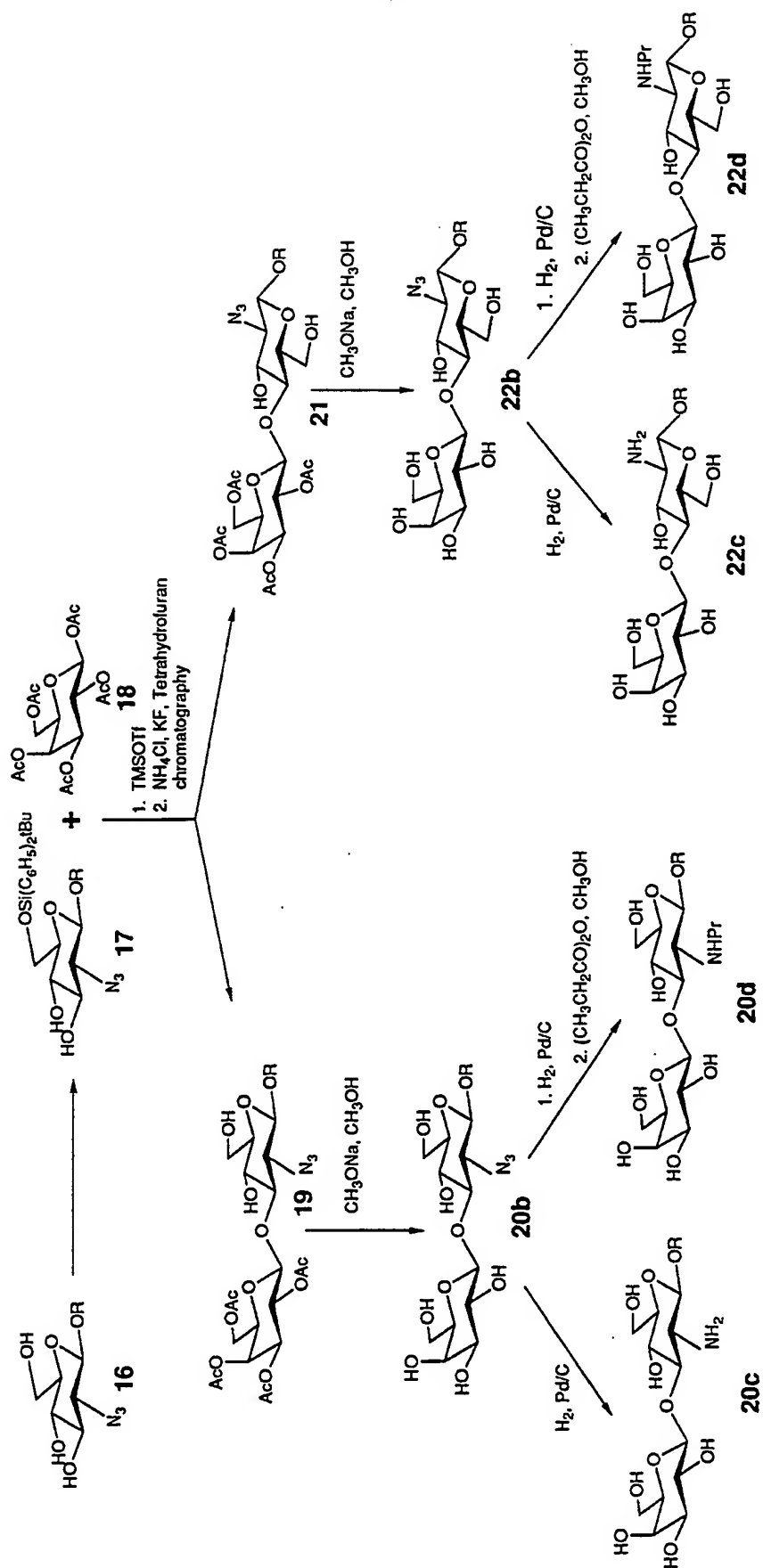

$$R = (CH_2)_8CO_2CH_3 \quad Pr = C(O)CH_2CH_3$$

Figure 5

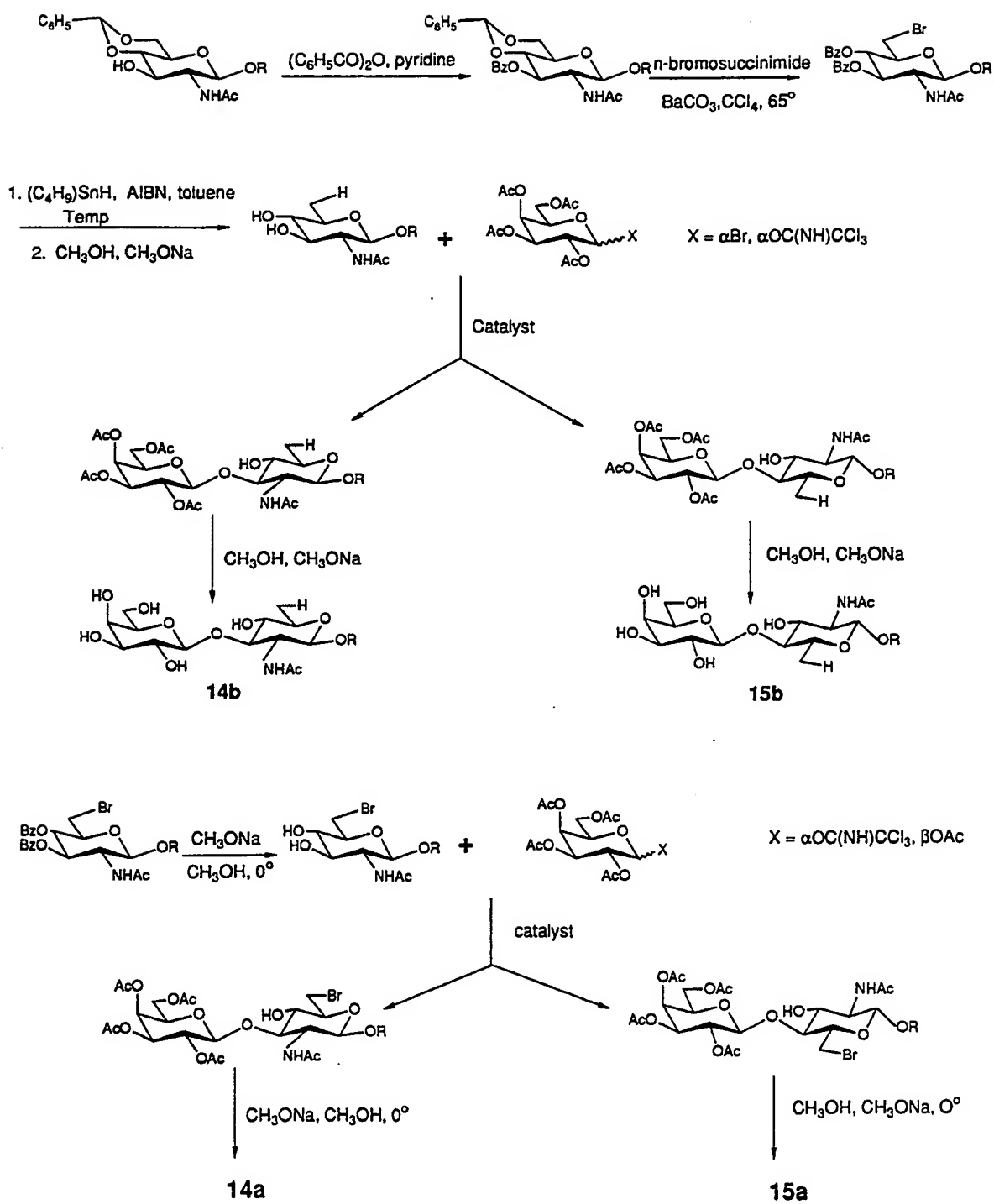


FIGURE 6

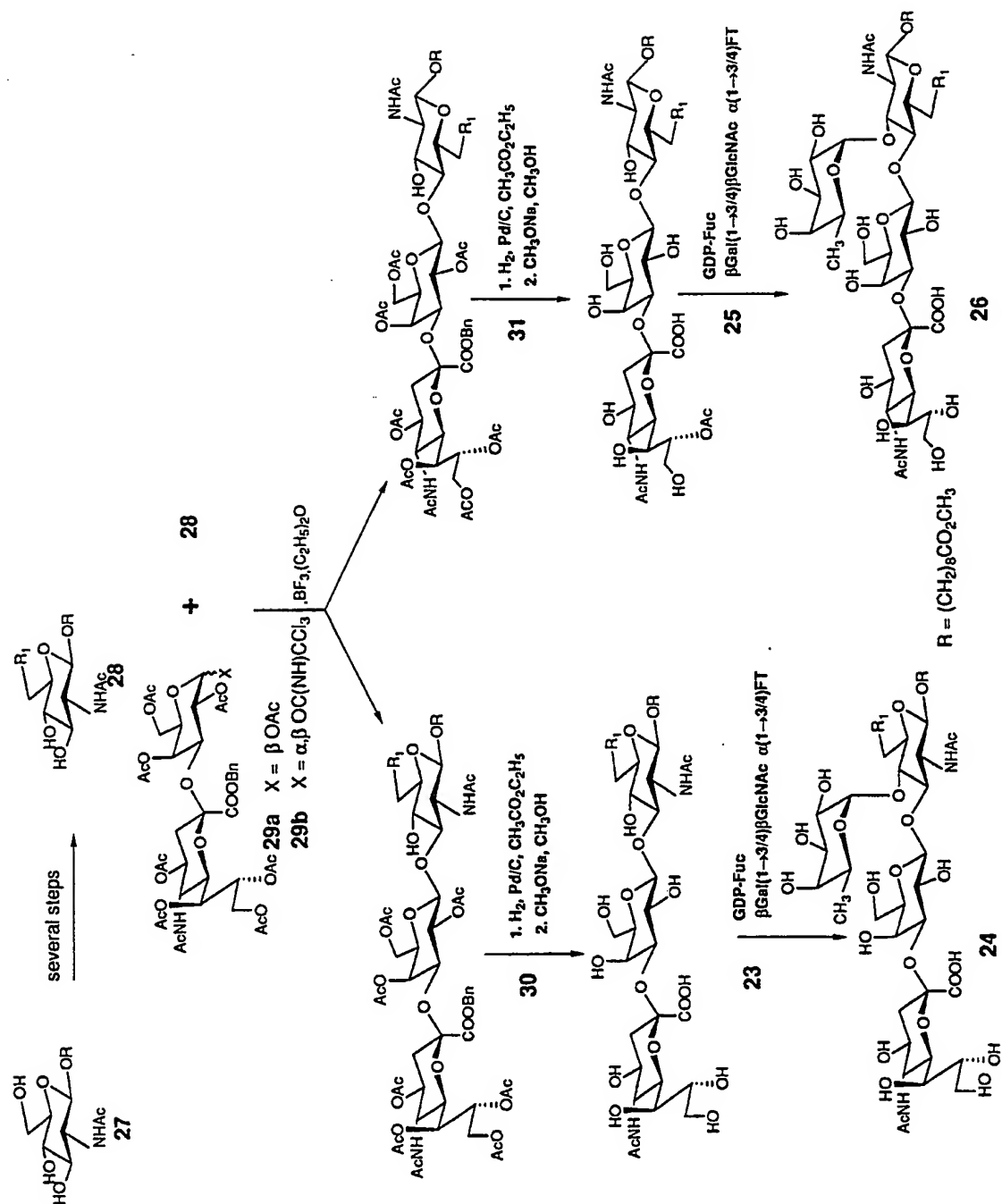


FIGURE 7

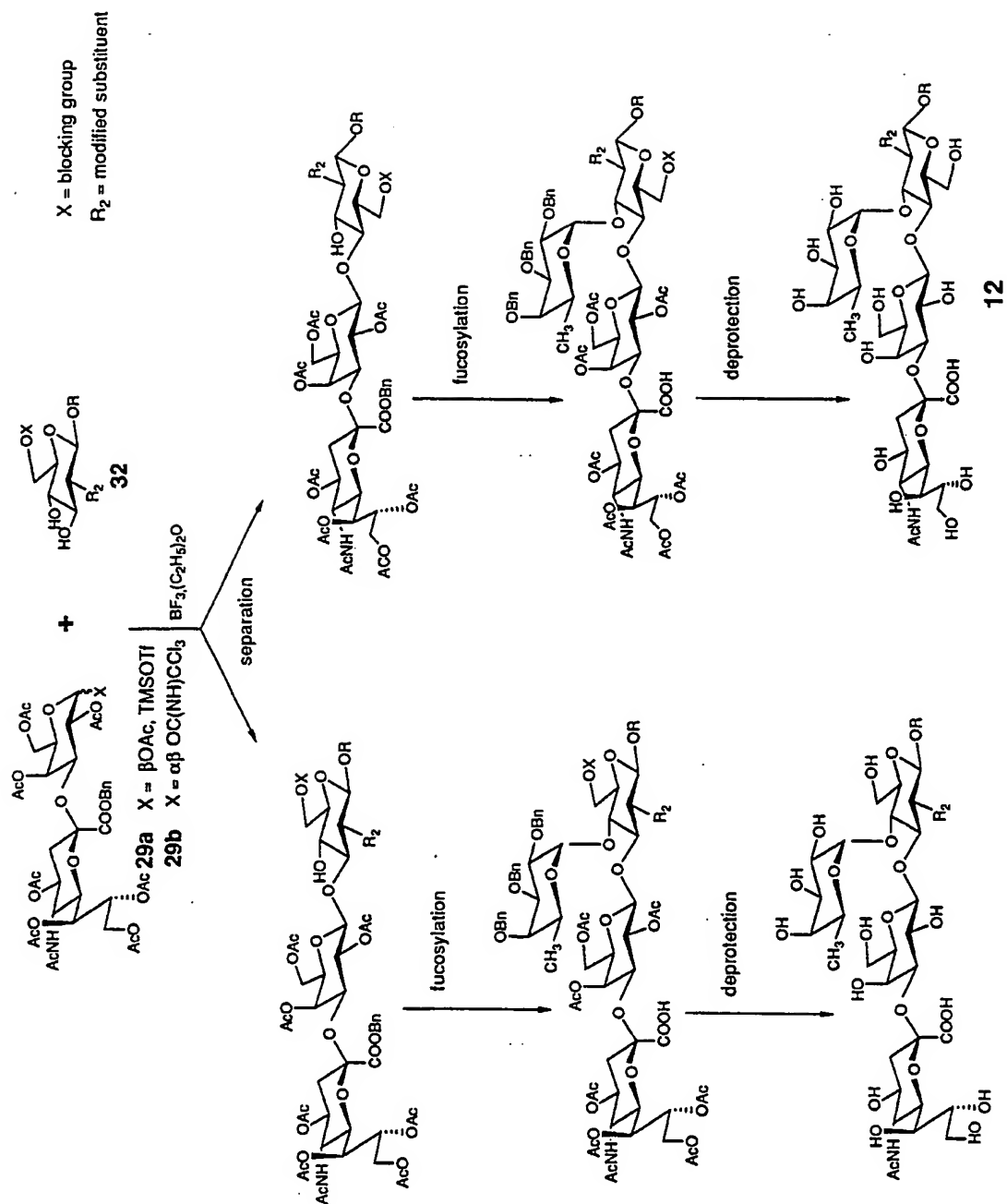


Figure 8

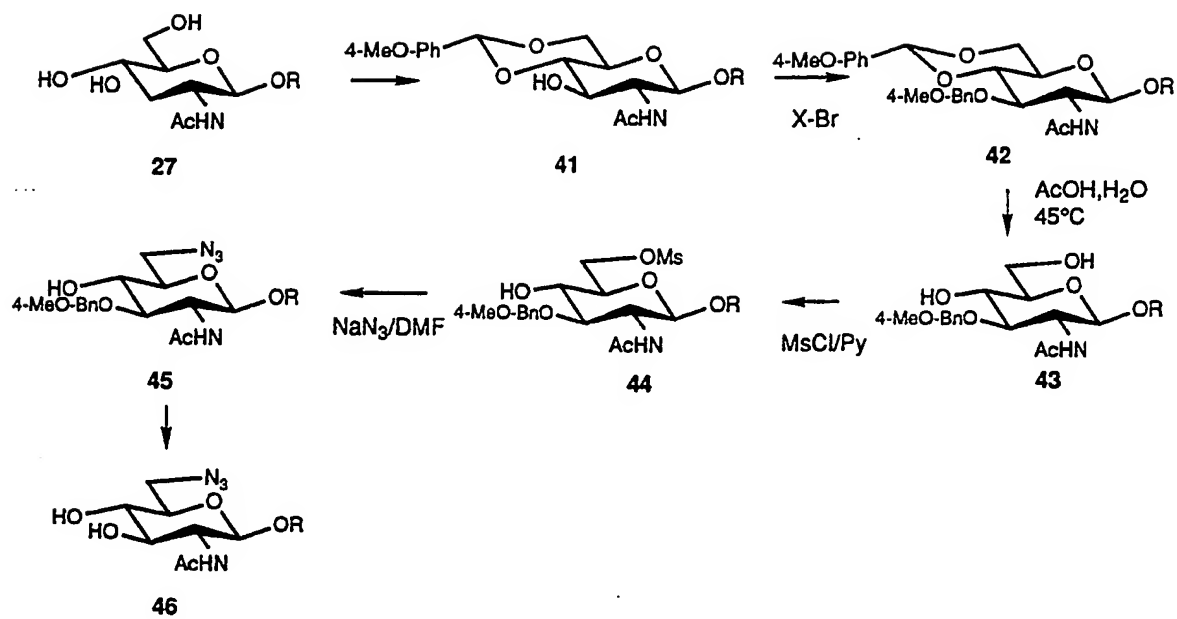
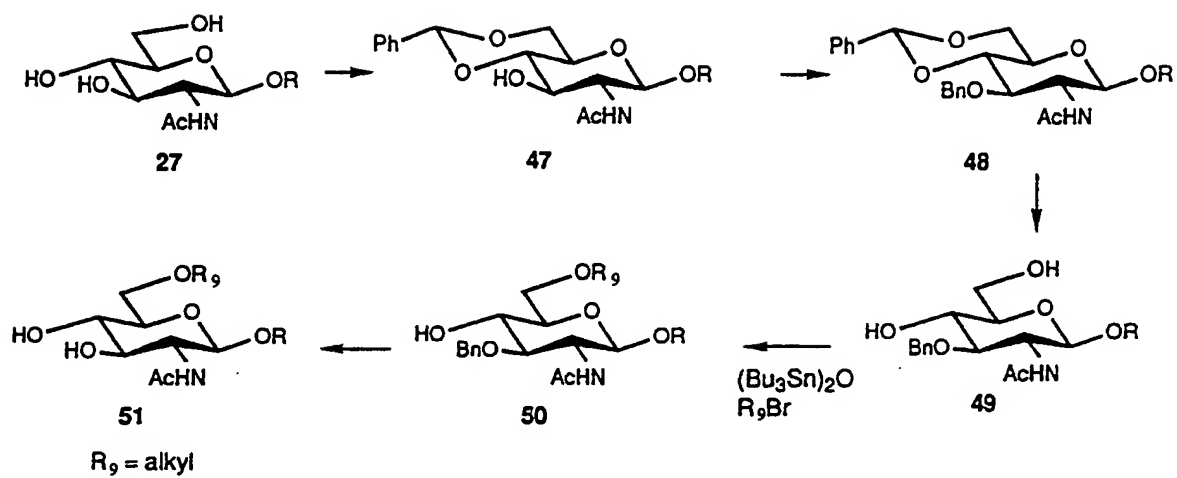


Figure 9



INTERNATIONAL SEARCH REPORT

PCT/CA 92/00246

International Application No

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ⁶		
According to International Patent Classification (IPC) or to both National Classification and IPC		
Int.Cl. 5 C07H3/06; C12N9/10;	C07H15/04; A61K31/70;	C12P19/26; A61K39/385 C12P19/44
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁷		
Classification System	Classification Symbols	
Int.Cl. 5	C07H ; C12P ; A61K	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁸		
III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹		
Category ¹⁰	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
A	CARBOHYDRATE RESEARCH. vol. 190, no. 1, 1989, AMSTERDAM NL pages 1 - 11 PALCIC, MONICA M. ET AL. 'Enzymic synthesis of oligosaccharides terminating in the tumor-associated sialyl-Lewis-a determinant' cited in the application see page 1 - page 7 ---	1-11, 14
X	EP,A,0 319 253 (CHEMBIOMED, LTD.) 7 June 1989	15
A	see page 8, line 30 - page 10, line 11; claims 1-3,5; figure 6 & US,A,5 079 353 cited in the application ---	12, 13
-/--		
<p>¹⁰ Special categories of cited documents:</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</p> <p>"&" document member of the same patent family</p>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search 28 SEPTEMBER 1992		Date of Mailing of this International Search Report 10 4. 10. 92
International Searching Authority EUROPEAN PATENT OFFICE		Signature of Authorized Officer DAY G.J.

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category °	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
A	<p>CARBOHYDRATE RESEARCH. vol. 174, 1988, AMSTERDAM NL pages 211 - 237 SPOHR. U. AND LEMIEUX R.U. 'SYNTHETIC, CONFORMATIONAL, AND IMMUNOCHEMICAL STUDIES OF MODIFIED LEWIS b AND Y HUMAN BLOOD-GROUP DETERMINANTS TO SERVE AS PROBES FOR THE COMBINING SITE OF THE LECTIN IV OF Griffonia simplicifolia' * Schemes 1 and 2 *</p> <p>-----</p>	1

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For more details about this annex : see Official Journal of the European Patent Office, No. 12/82